

Synthesis of 2',3'-Didehydro-2',3'-dideoxynucleosides by Reaction of 5'-Protected Nucleoside 2',3'-Dimesylates with Telluride Dianion: A General Route from *Cis* Vicinal Diols to Olefins

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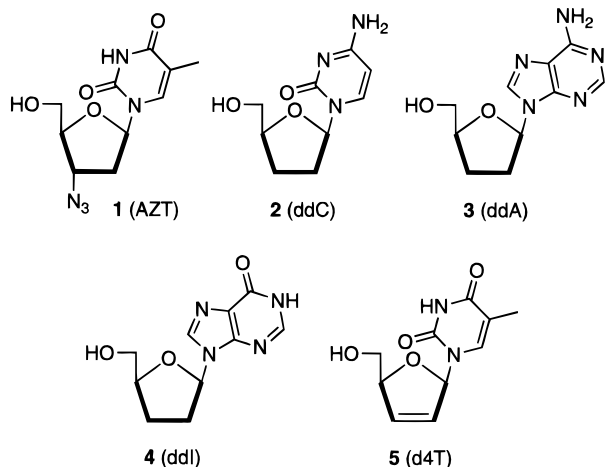
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2',3'-Dimesylates of 5'-protected nucleosides are converted into the corresponding 2',3'-didehydro-2',3'-dideoxy compounds by treatment with telluride dianion in the form of the sodium or lithium salt. The method is well-suited to the preparation of unsaturated nucleosides that can be converted into compounds that are believed to be useful in the treatment of AIDS. The deoxygenation is general for vicinal dimesylates that have, or may adopt, a synperiplanar conformation. With straight chain compounds the reaction is stereospecific. In some cases, similar, but slower, deoxygenations can be performed with selenide dianion.

Introduction

Replication of the human immunodeficiency virus is disrupted by certain deoxy nucleosides,¹ and several compounds of this type are currently important because of their potential, or actual, merit in the treatment of AIDS. The best known of these compounds² are 3'-azido-3'-deoxythymidine (**1**, AZT), 2',3'-dideoxycytidine (ddC, **2**), 2',3'-dideoxyadenosine (ddA, **3**), 2',3'-dideoxyinosine (ddI, **4**), and 2',3'-didehydro-3'-deoxythymidine (d4T, **5**), and the development of synthetic routes to these, and related, materials has become a prominent subject^{3,4} in medicinal chemistry.



2',3'-Dideoxynucleosides are often made from suitably protected nucleosides, and at least two approaches are used: (a) replacement of hydroxyl groups by hydrogen,⁵

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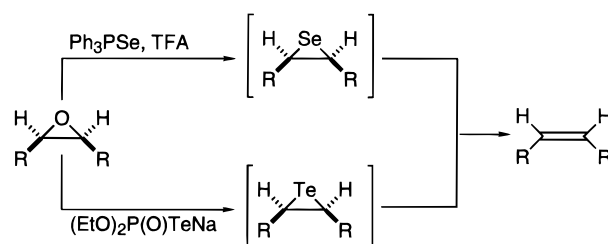
(2) (a) FDA approval has been granted for AZT, DDC, DDI, and d4T: *Chem. Eng. News* **1994**, *72*(27), 22 (July 4, 1994). (b) DDI is made from DDA by enzymatic replacement of the NH₂ by OH: Beach, C. M.; Evans, R. K.; Coleman, M. S. *Nucleosides Nucleotides* **1991**, *10*, 1499.

(3) For a review on AIDS-driven nucleoside chemistry: Huryn, D. M.; Okabe, M. *Chem. Rev.* **1992**, *92*, 1745.

(4) Dueholm, K. L.; Pedersen, E. B. *Synthesis* **1992**, 1.

(5) In the case of the thymidine series, only 3'-deoxygenation would, of course, be needed.

Scheme 1



and (b) double deoxygenation to the 2',3'-didehydro-2',3'-dideoxy compound, followed by hydrogenation. Both routes have been examined extensively,^{3,6-8} but the development of new methodology is warranted⁹ in view of the medical and commercial importance of the area. We report full details¹⁰ of an approach based on tellurium chemistry.

The starting point for this work was our experience in converting epoxides into olefins by treatment with triphenylphosphine selenide in the presence of acid (Scheme 1)¹¹ or by treatment with sodium diethyl phosphorotel-

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(7) For more recent developments, see: (a) Liu, Z.; Classon, B.; Samuelsson, B. *J. Org. Chem.* **1990**, *55*, 4273 and references therein. (b) Luzzio, F. A.; Menes, M. E. *J. Org. Chem.* **1994**, *59*, 7267.

(8) For recent examples of the preparation of 2',3'-dideoxynucleosides from nucleosides, see: (a) Dunkel, M.; Pfeleiderer, W. *Nucleosides Nucleotides* **1992**, *11*, 787. (b) Seela, F.; Muth, H.-P.; Bindig, U. *Synthesis* **1988**, 670. (c) Robins, M. J.; Hansske, F.; Low, N. H.; Park, J. I. *Tetrahedron Lett.* **1984**, *25*, 367. (d) Talekar, R. R.; Coe, P. L.; Walker, R. T. *Synthesis* **1993**, 303, and references therein. (e) Nair, V. *Nucleosides Nucleotides* **1989**, *8*, 699. (f) Prisbe, E. J.; Martin, J. C. *Synth. Commun.* **1985**, *15*, 401. (g) Lin, T.-S.; Yang, J.-H.; Liu, M.-C.; Zhu, J.-L. *Tetrahedron Lett.* **1990**, *31*, 3829. (h) Yamaguchi, T.; Saneyoshi, M. *Nucleosides Nucleotides* **1992**, *11*, 373. (i) Dorland, E.; Serafinowski, P. *Synthesis* **1992**, 477. (j) Chu, C. K.; Bhadti, V. S.; Doboszewski, B.; Gu, Z. P.; Kosugi, Y.; Pullaiah, K. C.; Van Roey, P. *J. Org. Chem.* **1989**, *54*, 2217. (k) Barton, D. H. R.; Jang, D. O.; Jaszberenyi, J. Cs. *Tetrahedron Lett.* **1991**, *32*, 7187. (l) Chen, B.-C.; Quinlan, S. L.; Stark, D. R.; Reid, J. G.; Audia, V. H.; George, J. G.; Eisenreich, E.; Brundidge, S. P.; Racha, S.; Spector, R. H. *Tetrahedron Lett.* **1995**, *36*, 7957.

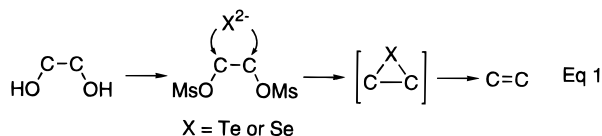
(9) For an evaluation of some of the classical methods, see: (a) References 8j. (b) Mansuri, M. M.; Starrett, J. E., Jr.; Wos, J. A.; Tortolani, D. R.; Brodfuehrer, P. R.; Howell, H. G.; Martin, J. C. *J. Org. Chem.* **1989**, *54*, 4780.

(10) Preliminary communication: Clive, D. L. J.; Wickens, P. L. *J. Chem. Soc., Chem. Commun.* **1993**, 923.

(11) Clive, D. L. J.; Denyer, C. V. *J. Chem. Soc., Chem. Commun.* **1973**, 253.

(12) Clive, D. L. J.; Menchen, S. M. *J. Org. Chem.* **1980**, *45*, 2347.

luroate (Scheme 1).¹² In each case, we interpreted the course of the reaction in terms of a three-membered ring intermediate, which spontaneously expelled the heavy atom.^{13,14} With this as background, we wondered if vicinal dimesylates might also be converted, transiently, into epitellurides or episelenides and then into olefins by the action of Na₂Te or Na₂Se, respectively (eq 1).



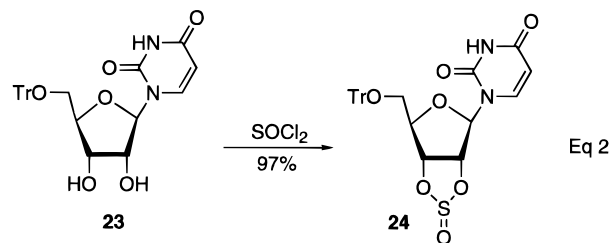
Vicinal dibromides have been converted into olefins by reaction with telluride¹⁵ or selenide¹⁶ dianions, but the use of dimesylates would be more readily applicable to ribonucleosides by direct esterification of the hydroxyl groups. In the event, the reaction summarized by eq 1 was easily reduced to practice, especially for X = Te, and our results are collected in Table 1; the method is mild, efficient, and general. We examined first the naphthalene derivative **6** (Table 1, entry 1), and then we made a survey of the main variables.

Use of Dimesylates. Our procedure requires that each of the vicinal hydroxyl groups be converted into a leaving group of such a type that only the original hydroxyl-bearing carbon—and not the other oxygen attachment—be susceptible to nucleophilic attack. This requirement is normally satisfied by sulfonate esters, and from the outset, we used dimesylates, which are simple and cheap to prepare.¹⁷ Yields in our mesylations were generally above 80%.

In one case (Table 1, entry 12) we also examined a ditosylate, but could detect no advantage, and in fact, the yield was lower in both steps than for the corresponding dimesylate.

Cyclic sulfates were also considered,^{18,19} but we found these compounds difficult to make (by oxidation of the corresponding sulfites)—at least in the few examples that we tried.²⁰ Cyclic sulfites,²¹ in contrast, are easy to prepare (e.g., eq 2) but, at least with **24** as a test case,

those do not give olefins under our standard conditions, the starting diol being recovered.^{19b,22}



Simple vicinal dimesylates have been converted into olefins by the action of sodium naphthalenide²³ or of iodide ion,²⁴ and so we tested both of these systems on the dimesylate **9**. With sodium naphthalenide, a complex mixture resulted, and with NaI/18-crown-6/Zn dust or NaI/18-crown-6/Zn–Cu couple (in DMF) we did not detect formation of the olefin.

Source of Telluride Dianion. Many of our experiments were carried out on a small scale, and for reasons of convenience we generated the telluride dianion by addition of Et₃BHLi (Super-Hydride) to metallic Te.²⁵ The Et₃B liberated in these reactions does not seem to play a significant part¹³ in the deoxygenation, because the same results are obtained by generating the telluride dianion from Na and Te in liquid NH₃—a process, of course, that affords reagent free of Lewis acids.

We also evaluated several other known, and potential, ways of generating the telluride dianion, and some of these are listed in Table 2. Apart from the Te/Et₃BHLi and Na/Te/liquid NH₃ systems of Table 1 and Te/*s*-Bu₃BHLi (Table 2), none of the methods was successful, and in many cases we formed the impression that the presence of base led to destruction of the starting material and/or product.

In the small-scale experiments summarized in Table 2, we sometimes found that an excess of reducing agent was needed to dissolve all the Te, and in these cases we often obtained a poor yield. For this reason, we sought to quench any excess of hydride (in an experiment with Et₃BHLi) by addition of acetone (Table 2, entry 11) or of EtOH (Table 2, entry 12), but any potential advantage was offset by the (deleterious) generation of alkoxide.

In our hands, several of the reported preparations of telluride dianion gave material that was probably con-

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Table 1^a

Dimesylate (Yield from diol)		Olefin	Dimesylate (Yield from diol)		Olefin
(1)			(15)		
	$\xrightarrow[\text{Rt / 20 h}]{\text{Li}_2\text{Te/THF}}$	(88%)		$\xrightarrow[\text{Rt / 96 h}]{\text{Li}_2\text{Te/THF}}$	(14%) ^d
(2)			(16)		
	$\xrightarrow[\text{Bath 100 }^\circ\text{C / 2 h}]{\text{Li}_2\text{Te/THF}}$	(83%)		$\xrightarrow[\text{Rt / 20 h}]{\text{Li}_2\text{Te/THF}}$	(78%)
(3)			(17)		
	$\xrightarrow[\text{Bath 100 }^\circ\text{C / 4 h}]{\text{Li}_2\text{Se/THF-Dioxane}}$	(76%)		$\xrightarrow[\text{Rt / 16 h}]{\text{Li}_2\text{Te/THF}}$	(ca. 89%)
(4)			(18)		
	$\xrightarrow[\text{Bath 105 }^\circ\text{C / 20 h}]{\text{Li}_2\text{Te/THF-Dioxane}}$	(69%)		$\xrightarrow[\text{THF / 40 }^\circ\text{C / 24 h}]{2.5 \text{ equiv Li}_2\text{Te}}$	(ca. 93%) ^{f,i}
(5)			(19)		
	$\xrightarrow[\text{Rt / 20 h}]{\text{Na}_2\text{Te/THF}}$	(75-93%)		$\xrightarrow[\text{Rt / 12 h}]{\text{Li}_2\text{Te/THF}}$	(67%) ^k
(6)			(20)		
	$\xrightarrow[\text{Rt / 48 h}]{\text{Li}_2\text{Te/THF-Dioxane}}$	(80%)		$\xrightarrow[\text{Rt / 17 h}]{\text{Li}_2\text{Te/THF}}$	(75%) ^k
(7)			(21)		Complex mixture
	$\xrightarrow[\text{Rt / 48 h}]{\text{Na}_2\text{Se/THF}}$	(51%)		$\xrightarrow[\text{Rt / 12 h}]{\text{Li}_2\text{Te/THF}}$	Complex mixture
(8)			(22)		Complex mixture
	$\xrightarrow[\text{Rt / 20 h}]{\text{Li}_2\text{Se/THF}}$	(65%)		$\xrightarrow[\text{Rt / 12 h}]{\text{Li}_2\text{Te/THF}}$	Complex mixture
(9)			(23)		No reaction
	$\xrightarrow[\text{Rt / 16 h}]{\text{Li}_2\text{Te/THF/MeCN}^b}$	(99%)		$\xrightarrow[\text{Rt / 12 h}]{\text{Li}_2\text{Te/THF}}$	No reaction
(10)			(24)		No reaction. Starting material recovered in 70% yield
	$\xrightarrow[\text{Rt / 24 h}]{\text{Na}_2\text{Te/MeCN}}$	(44%)			
(11)					
	$\xrightarrow[\text{Rt / 48 h}]{\text{Li}_2\text{Te / THF}}$	(90%)			
(12)					
	$\xrightarrow[\text{Rt / 24 h}]{\text{Li}_2\text{Te / THF}}$	(60%)			
(13)					
	$\xrightarrow[\text{Rt / 14 h}]{\text{Li}_2\text{Te/THF}}$	(83%)			
(14)					
	$\xrightarrow[\text{Rt / 24 h}]{\text{Na}_2\text{Te/THF}}$	(42%)			

^a All lithium salts were generated using Et_3BHLi , and sodium salts were prepared from Na and Te (or Se) in liquid NH_3 , followed by evaporation of the NH_3 . ^b MeCN was added after formation of Li_2Te . ^c Hydrogenation gave the saturated compound (61%). ^d Possibly, the poor yield is due, in part, to reaction of the excess of Et_3BHLi with the acetate group. ^e X = N=CH(NMe₂). ^f DMT = dimethoxytrityl. ^g Hydrogenation and removal of the N-protecting group gave the saturated product in 43% overall yield. ^h Yield not optimized; recycling as previously described for analogous compounds (Kawana, M.; Kuzuhara, H. *J. Chem. Soc., Perkin Trans. 1* **1992**, 469) was not done. ⁱ Trace impurities are present (¹H NMR). ^j Reaction is fast; no starting material remains after 1 h. ^k Yield not optimized.

taminated with polytellurides, as suggested by the generation of an intense dark red or purple color.²⁶ *Dilithium ditelluride*²⁵ itself does not lead to olefinic material, at least as judged by attempted reaction with dimesylate **9**. Sodium hydrogen telluride²⁷ is likewise unsuccessful (with the same compound).

We also tried several electrochemical methods²⁹ for generating the telluride dianion, but yields of dihydrodideoxynucleosides were very poor. Only with the

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Table 2

entry	reaction mixture	substrate	remarks
1	Te/sodium naphthalenide ^{28a}	9	none of desired olefin
2	Te/Rongalite/aq NaOH ^{28c,d}	9	none of desired olefin
3	Te/Rongalite/DMF/NaHCO ₃	9	none of desired olefin
4	Te/hydrazine/NaOH ^{28e}	9	none of desired olefin
5	Te/hydrazine/LiOH/DMF or THF	none	no reduction of Te
6	Te/NaBH ₄ /THF or DMF ^{28f-h}	9	none of desired olefin
7	Te/Bu ₄ NBH ₄ ^{28d}	9	none of desired olefin
8	Te/LiBH ₄ /DMF	none	no reduction of Te
9	Te/Red-Al/THF	9	desired olefin (6%)
10	Te/LiAlH ₄ /THF	9	no reduction of Te
11	Te/Et ₃ BHLi; addition of acetone	9	desired olefin (30%)
12	Te/Et ₃ BHLi; addition of EtOH	9	desired olefin (38%)
13	Te/DIBAL-H/THF	none	no reduction of Te
14	Te/NaH/DMF or dioxane ^{28i-k}	9	none of desired olefin
15	Te/LiH/THF	9	no reduction of Te
16	Te/LiH/Et ₃ B/THF	9	no reduction of Te
17	Se/LiH/THF	9	no reduction of Se
18	Te/ <i>s</i> -Bu ₃ BHLi	9	desired olefin (62%)

simple dimesylate **6** could the desired olefin be isolated (43% yield).

In our standard deoxygenation with Te²⁻, the Te precipitates at the end of the experiment. In one small-scale test (0.128 mmol of dimesylate **15**), we recovered 53% of the original amount of Te by filtering the crude reaction mixture through a bed of β -cyclodextrin and then dissolving the cyclodextrin in boiling water. Filtration of the hot mixture afforded the Te as a fine powder. In this experiment the initial olefin was processed further: hydrogenation of the double bond and removal of the *N*-protecting group gave 5'-*O*-[bis(4-methoxyphenyl)phenylmethyl]-2',3'-dideoxyadenosine in 43% yield (over three steps).

Use of Selenide or Sulfide Dianion. Treatment of dimesylate **9** (Table 1, entries 7 and 8) with selenide dianion (from Se and Na/liquid NH₃³⁰ or Se and Et₃BHLi³¹) does lead to the expected olefin, but the reaction is slower than in the case of Te. An attempt (with **9**) to use sulfide dianion (from S and Et₃BHLi³²), in the hope of forming an episulfide, gave a complex mixture.

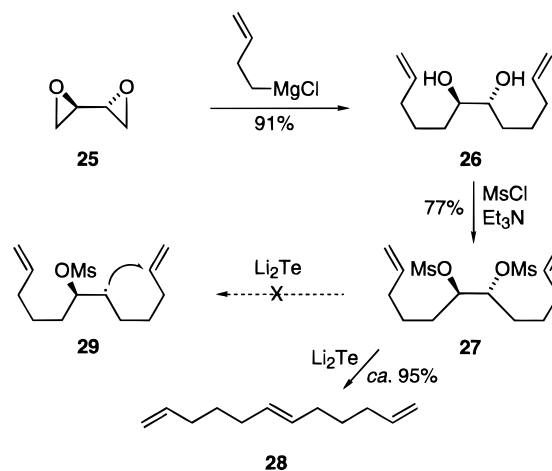
Effect of Solvent and Temperature. We generally used THF, as this solvent is inert to Et₃BHLi. However, when preformed Na₂Te (from Na, Te, and liquid NH₃) is used, either THF or MeCN is suitable. We did not make a careful comparison of yields in the two solvents; we simply established (Table 1, entries 9 and 10) that the desired olefin is obtained in acetonitrile.

We generally ran our experiments at room temperature, and although the reaction is slow, yields are often high (see Table 1).

Nature of the Dimesylate. As shown in Table 1, both terminal and internal acyclic dimesylates undergo the present reaction. In the case of dimesylates derived from cyclic diols, those in the nucleoside series work well—fortunately—but the sugar-derived substrate **22** is not suitable, and it is clear that the hydroxyl groups have to be *syn* coplanar (or close to that). This preferred arrangement is, of course, readily accessible to the acyclic compounds.

The reaction is sensitive to steric factors, and the dimesylate derived from the [2.2.1] bicyclic substrate **21** (Table 1, entry 23) is not converted into olefin—the

Scheme 2



material is simply recovered. The cyclooctane substrates^{23a,33} (Table 1, entries 21 and 22) gave complex mixtures.

Of the protecting groups used for the 5'-oxygen, only acetyl appears to be unsuitable (Table 1, entry 15).

Mechanistic Considerations. The stereochemical result with the open chain dimesylates **7** and **18**, i.e., the formation of *Z* and *E* olefins, respectively (Table 1, entries 2 and 20), is consistent with a simple nucleophilic displacement mechanism (eq 1), as is the sensitivity of the reaction to steric factors (Table 1, entries 21–23). However, our observations do not exclude a single electron transfer pathway. In an attempt to intercept possible carbon radical intermediates, arising from a SET mechanism, we prepared the dimesylate **27**, as shown in Scheme 2. Under our standard conditions, we were unable to detect any cyclized product and obtained only the olefin **28** (ca. 95% yield), whose stereochemistry was established³⁴ by NMR techniques. Our working hypothesis is, therefore, the simple double nucleophilic pathway.

(30) Durst, T.; Tin, K.-C. *Can. J. Chem.* **1970**, *48*, 845.

(34) The *E*-stereochemistry of the C(6)–C(7) double bond was determined from the value (16 Hz) of the coupling constant of the hydrogens on the double bond. A value of ca. 8–10 Hz would be observed if the stereochemistry were *Z*. The measurement was made by first decoupling the hydrogens on the C(5) and C(8) methylene groups to simplify the spectrum. The coupling constant was then measured from the proton satellites corresponding to R¹³CH=CH¹²CHR. In this situation (i.e., one carbon is ¹³C and the other is ¹²C) the molecule is unsymmetrical and the coupling can be measured.

(30) Cf. Rossi, R. A.; Peññory, A. B. *J. Org. Chem.* **1981**, *46*, 4580.
(31) Gladysz, J. A.; Hornby, J. L.; Garbe, J. E. *J. Org. Chem.* **1978**, *43*, 1204.

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Conclusion

Conversion of vicinal diols into the corresponding dimesylates, followed by treatment with Na₂Te (from Na and Te in liquid NH₃) or Li₂Te (from Te and Et₃BHLi), results in smooth deoxygenation to the olefin in the case of acyclic compounds or nucleosides. With nucleosides, the 5'-protecting group can be removed,^{9b,35,36} and hydrogenation³⁷ of the double bond gives the corresponding dideoxy compounds. Amino groups in the nucleoside base components do not necessarily have to be protected, and in general, the deoxygenation described here is well-suited to the preparation of deoxynucleosides.

Experimental Section

General Procedures. Unless stated to the contrary, the following conditions apply: Reactions were carried out under a slight static pressure of Ar that had been purified by passage through a column (3.5 × 42 cm) of R-311 catalyst³⁸ and then through a similar column of Drierite. Glassware was dried in an oven for at least 3 h before use (120 °C) and either cooled in a desiccator over Drierite or assembled quickly, sealed with rubber septa, and allowed to cool under a slight static pressure of Ar. Reaction mixtures were stirred by Teflon-coated magnetic stirring bars.

Hexane used for chromatography was distilled before use.

Products were isolated from solution by evaporation under water-aspirator vacuum at, or below, room temperature, using a rotary evaporator.

Cannula transfers were done under slight pressure (Ar), not by suction.

Commercial thin layer chromatography (TLC) plates (silica gel, Merck 60F-254) were used. Spots were detected by spraying the plate with a solution of phosphomolybdic acid³⁹ or *p*-anisaldehyde,⁴⁰ followed by charring with a heat gun or by examination under UV light. Silica gel for flash chromatography was Merck type 60 (230–400 mesh).

Dry solvents were prepared under an inert atmosphere and transferred by syringe or cannula. Dry THF and Et₂O were distilled from sodium and benzophenone ketyl. MeCN and pyridine were distilled from CaH₂.

FTIR measurements were made as casts from the specified solvent using potassium bromide plates.

The symbols *s*', *d*', *t*', and *q*' used for ¹³C NMR signals indicate zero, one, two, or three attached hydrogens, respectively.

Preparation of Sodium Telluride (Na₂Te).⁴¹ A 500-mL three-neck round-bottomed flask was charged with Te powder (200 mesh, 2.205 g, 17.28 mmol) and a stirring bar. Na (794 mg, 34.5 mmol) was placed in a sidearm addition tube, and the central neck of the flask was fitted with a condenser charged with dry ice/acetone and closed by a septum carrying both an entry needle for Ar and an exit needle leading to an oil bubbler. The third neck of the flask was temporarily closed by a septum, and the flask was flushed with Ar. The septum in the third neck was removed and immediately replaced by an adaptor (fitted with a tap) connected to a tank of liquid NH₃. The flask was then cooled with dry ice/MeCN, and NH₃ was led in until *ca.* 200 mL had collected. The ammonia inlet

was closed, and a slow stream of Ar was maintained. The stirrer was started, and the Na was added portionwise by tapping the sidearm addition tube. The mixture changed from red to bluish-green to white, by which stage formation of Na₂Te was complete. The cooling bath was removed, and stirring was continued overnight, during which time the coolant in the condenser attained room temperature and the NH₃ evaporated. The resulting beige Na₂Te (*ca.* 100% yield) was transferred in an Ar-filled glovebag to a storage flask.

The experiment was also done on a larger scale (Te, 6.096 g) with the same result.

Preparation of Sodium Selenide (Na₂Se).³⁰ A 500-mL three-neck round-bottomed flask was charged with Se powder (325 mesh, 2.847 g, 36.06 mmol) and a stirring bar. Na (1.741 g, 75.73 mmol) was placed in a sidearm addition tube, and the central neck of the flask was fitted with a condenser charged with dry ice/acetone and closed by a septum carrying both an entry needle for Ar and an exit needle leading to an oil bubbler. The third neck of the flask was temporarily closed by a septum, and the flask was flushed with Ar. The septum in the third neck was removed and immediately replaced by an adaptor (fitted with a tap) connected to a flask containing a small amount (*ca.* 1 g) of Na. This latter flask was, in turn, connected to a tank of liquid NH₃ and was then cooled with dry ice/MeCN. NH₃ was led in until *ca.* 200 mL had collected. The cooling bath was removed, and the NH₃ was distilled into the reaction vessel, which was cooled with dry ice/MeCN. The ammonia inlet was removed, and a slow stream of Ar was maintained. The stirrer was started, and the Na was added portionwise by tapping the sidearm addition tube. The mixture changed color and eventually became white, by which stage formation of Na₂Se was complete. The cooling bath was removed, and stirring was continued overnight, during which time the coolant in the condenser attained room temperature and the NH₃ evaporated. The resulting slightly orange Na₂Se (*ca.* 100% yield) was transferred in an Ar-filled glovebag to a storage flask.

4-(1-Naphthyl)butane-1,2-diol. OsO₄ (1.7 mL, 2.5% w/w solution of OsO₄ in *t*-BuOH) was added to a stirred solution of 1-(3-butenyl)naphthalene⁴² (**6a**) (1.121 g, 6.153 mmol) and 4-methylmorpholine *N*-oxide (1.033 g, 7.652 mmol) in acetone (30 mL) and water (15 mL). Stirring at room temperature was continued for 43 h. EtOAc (100 mL) was then added, and the organic layer was washed with water (1 × 100 mL) and aqueous Na₂SO₃ (10% w/v, 2 × 50 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (5 × 20 cm), using 7:3 EtOAc–hexane, gave 4-(1-naphthyl)butane-1,2-diol (1.200 g, 90%) as a colorless oil: FTIR (CH₂Cl₂ cast) 3360 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz) δ 1.70–2.05 (m, 4 H), 3.05–3.45 (m, 2 H), 3.50 (dd, *J* = 11.0, 7.0 Hz, 1 H), 3.71 (dd, *J* = 11.0, 3.0 Hz, 1 H), 3.75–3.95 (m, 1 H), 7.30–7.60 (m, 4 H), 7.60–7.80 (m, 1 H), 7.80–7.95 (m, 1 H), 7.95–8.20 (m, 1 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 28.93, 34.02, 66.86, 71.89, 123.74, 125.55, 125.58, 125.92, 126.07, 126.82, 128.85, 131.81, 133.97, 137.89; exact mass *m/z* calcd for C₁₄H₁₆O₂ 216.1151, found 216.1151.

4-(1-Naphthyl)butane-1,2-diol Bis(methanesulfonate) (6). MeSO₂Cl (1.6 mL, 20 mmol) in CH₂Cl₂ (10 mL) was added dropwise to a stirred and cooled (0 °C) solution of 4-(1-naphthyl)butane-1,2-diol (1.123 g, 5.192 mmol) and pyridine (3.4 mL, 41 mmol) in CH₂Cl₂ (10 mL). The ice bath was removed, and stirring was continued for 16 h. The mixture was poured onto ice (*ca.* 50 g) and extracted with EtOAc (1 × 100 mL). The organic extract was washed with aqueous CuSO₄ (10% w/v, 2 × 50 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (3 × 30 cm), using 1:1 EtOAc–hexane, gave **6** (1.849 g, 96%) as a colorless oil: FTIR (CH₂Cl₂ cast) 1356, 1173 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz) δ 2.05–2.40 (m, 2 H), 3.0–3.10 (s, 3 H), 3.10–3.15 (s, 3 H), 3.15–3.45 (m, 2 H), 4.30 (dd, *J* = 11.0, 6.0 Hz, 1 H), 4.44 (dd, *J* = 11.0, 3.0 Hz, 1 H), 4.95–5.10 (m, 1 H),

(35) (a) Cosford, N. D. P.; Schinazi, R. F. *J. Org. Chem.* **1991**, *56*, 2161. (b) Cosford, N. D. P.; Schinazi, R. F. *Nucleosides Nucleotides* **1993**, *12*, 149.

(36) E.g. (a) Kaskar, B.; Markovac, A. *J. Heterocycl. Chem.* **1989**, *26*, 1531. Removal of 5'-*O*-trityl and acyl groups from 3'-deoxythymidine: (b) Sekine, M.; Nakanishi, T. *J. Org. Chem.* **1990**, *55*, 924.

(37) Manchand, P. S.; Belica, P. S.; Holman, M. J.; Huang, T.-N.; Maehr, H.; Tam, S. Y.-K.; Yang, R. T. *J. Org. Chem.* **1992**, *57*, 3473.

(38) Supplied by Chemical Dynamics Corp., South Plainfield, NJ.

(39) Phosphomolybdic acid (15 g) and (NH₄)₂Ce(NO₃)₆ (2.5 g) dissolved in a mixture of water (485 mL) and concentrated H₂SO₄ (15 mL).

(40) *p*-Anisaldehyde (15 drops) was added to concentrated H₂SO₄ (6 mL) and EtOH (94 mL).

(41) Cf. Brauer, G., Ed. *Handbuch der Präparativen Anorganischen Chemie*; Ferdinand Enke: Stuttgart, 1975; Vol. I, p 431.

(42) Lambert, J. B.; Fabricius, D. M.; Hoard, J. A. *J. Org. Chem.* **1979**, *44*, 1480. In preparing the butenyl naphthalene, we used allyllithium, generated (Seyferth, D.; Weiner, M. A.; *J. Org. Chem.* **1961**, *26*, 4797) from triphenyl(2-propenyl)stannane.

7.30–7.65 (m, 4 H), 7.70–7.80 (m, 1 H), 7.80–7.93 (m, 1 H), 7.93–8.05 (m, 1 H); ^{13}C NMR (CDCl_3 , 75.5 MHz) δ 28.25, 32.08, 37.74, 38.90, 69.46, 78.66, 123.26, 125.63, 125.76, 126.30 (two overlapping signals), 127.40, 129.01, 131.49, 134.00, 135.99; exact mass m/z calcd for $\text{C}_{16}\text{H}_{20}\text{O}_6\text{S}_2$ 372.0702, found 372.0694.

General Procedure for Deoxygenation: 1-(3-Butenyl)naphthalene (6a) from Dimesylate 6. Te powder (200 mesh, 167 mg, 1.31 mmol) and a small stirring bar were placed in a dry round-bottomed flask fused onto a reflux condenser. The flask was closed with a septum and flushed with Ar. A solution of Et_3BHLi (1 M in THF, 3.4 mL, 3.4 mmol) was injected, and the mixture was stirred until a milky white suspension had formed (ca. 5 h). 4-(1-Naphthyl)butane-1,2-diol bis(methanesulfonate) (**6**) (488 mg, 1.31 mmol) in THF (5 mL) was then injected dropwise, and the mixture was stirred for 20 h. The mixture was washed out of the flask with acetone and evaporated at room temperature. Flash chromatography of the residue over silica gel (2 \times 30 cm), using hexane, gave **6a** (210 mg, 88%), spectroscopically identical to an authentic sample.

Benzyl (9*RS*,10*SR*)-9,10-Dihydroxyoctadecanoate. (a) Oleic Acid Benzyl Ester (7a). The following procedure is different from that given in the literature.⁴³

BuOH (2.59 g, 24.0 mmol) was added to a stirred solution of oleic acid (5.70 g, 20.2 mmol) and $\text{TsOH} \cdot \text{H}_2\text{O}$ (200 mg, 0.6 mmol) in PhH (20 mL). The mixture was refluxed for 22 h using a Dean–Stark apparatus. Evaporation of the solvent and flash chromatography of the residue over silica gel (6 \times 40 cm), using first hexane and then 1:9 EtOAc –hexane, gave **7a** (7.55 g, 100%) as a colorless oil: FTIR (CH_2Cl_2 , cast) 2925, 2853, 1739 cm^{-1} ; ^1H NMR (CDCl_3 , 200 MHz) δ 0.80–0.98 (m, 3 H), 1.08–1.45 (m, 20 H), 1.52–1.75 (m, 2 H), 1.90–2.01 (m, 4 H), 2.30–2.42 (apparent t, $J = 8.0$ Hz, 2 H), 5.12 (apparent s, 2 H), 5.30–5.40 (m, 2 H), 7.30–7.42 (m, 5 H); ^{13}C NMR (CDCl_3 , 75.5 MHz) δ 14.13, 22.71, 24.98, 27.26, 29.02, 29.13, 29.17, 29.28, 29.36, 29.49, 29.56, 29.62, 29.80, 31.94, 34.36, 66.07, 128.17 (two overlapping signals), 128.55, 129.77, 130.02, 136.21, 173.64; mass (CI) m/z calcd for $\text{C}_{25}\text{H}_{40}\text{O}_2$ 372, found 372.

(b) Benzyl (9*RS*,10*SR*)-9,10-Dihydroxyoctadecanoate. OsO_4 (3.95 mL, 2.5% w/w solution of OsO_4 in $t\text{-BuOH}$) was added to a solution of oleic acid benzyl ester (5.658 g, 15.18 mmol) and 4-methylmorpholine *N*-oxide (3.749 g, 27.77 mmol) in acetone (500 mL) and water (38 mL). The mixture was stirred at room temperature for 24 h and then evaporated at room temperature to ca. 100 mL. EtOAc (200 mL) was added, and the organic layer was washed with water (1 \times 200 mL) and aqueous Na_2SO_3 (10% w/v, 3 \times 200 mL), dried (MgSO_4), and evaporated. Flash chromatography of the residue over silica gel (10 \times 50 cm), using 3.2:96.8 MeOH – CHCl_3 , gave benzyl (9*RS*,10*SR*)-9,10-dihydroxyoctadecanoate (4.432 g, 72%) as a colorless oil: FTIR (CHCl_3 , cast) 3280, 1735 cm^{-1} ; ^1H NMR (CDCl_3 , 400 MHz) δ 0.80–1.00 (m, 3 H), 1.20–1.58 (m, 24 H), 1.58–1.75 (m, 2 H), 1.95 (br s, 2 H), 2.35 (t, $J = 7.2$, 2 H), 3.50–3.70 (m, 2 H), 5.11 (s, 2 H), 7.28–7.45 (m, 5 H); ^{13}C NMR (CDCl_3 , 75.5 MHz) δ 14.13, 22.70, 24.93, 25.95, 26.06, 29.06, 29.18, 29.31, 29.46, 29.59, 29.73, 31.21, 31.30, 31.91, 34.34, 66.12, 74.69, 74.76, 128.20 (two overlapping signals), 128.58, 136.70, 173.70; exact mass m/z calcd for $\text{C}_{25}\text{H}_{38}\text{O}_2$ (M – 2 H_2O) 370.2873, found 370.2869; mass (CI) m/z calcd for $\text{C}_{25}\text{H}_{42}\text{O}_4$ 406, found 424 (M + 18).

Benzyl (9*RS*,10*SR*)-9,10-Dihydroxyoctadecanoate Bis(methanesulfonate) (7). MeSO_2Cl (1.8 mL, 23 mmol) in CHCl_3 (4 mL) was added dropwise to a stirred and cooled (0 $^\circ\text{C}$) solution of benzyl (9*RS*,10*SR*)-9,10-dihydroxyoctadecanoate (1.170 g, 2.878 mmol) and pyridine (3.80 mL, 46.0 mmol) in CHCl_3 (11 mL). The ice bath was removed, and stirring was continued for 40 h. The mixture was poured onto ice (ca. 50 g) and extracted with CHCl_3 (200 mL). The organic extract was washed with aqueous CuSO_4 (10% w/v, 2 \times 100 mL) and aqueous NaOH (0.5 M, 1 \times 50 mL), dried (MgSO_4), and evaporated. Flash chromatography of the residue over silica gel (3 \times 30 cm), using 1:19 MeOH – CHCl_3 , gave **7** (1.532

g, 95%), which contained a trace impurity (^1H NMR, 200 MHz, small signals at δ 3.0 and 3.1) but was suitable for the next stage: FTIR (CHCl_3 , cast) 1735, 1357, 1175 cm^{-1} ; ^1H NMR (CDCl_3 , 200 MHz) δ 0.78–0.98 (m, 3 H), 1.18–1.88 (m, 26 H), 2.35 (t, $J = 7.0$ Hz, 2 H), 3.09 (s, 6 H), 4.68–4.88 (m, 2 H), 5.10 (s, 2 H), 7.25–7.38 (m, 5 H); ^{13}C NMR (CDCl_3 , 75.5 MHz) δ 14.07, 22.50, 22.61, 24.81, 25.36, 25.45, 28.91 (three overlapping signals), 29.14, 29.27, 29.58, 29.65, 31.77, 34.22, 38.82 (two overlapping signals), 66.06, 82.81, 82.91, 128.15 (two overlapping signals), 128.54, 136.14, 173.52; exact mass m/z calcd for $\text{C}_{27}\text{H}_{47}\text{O}_8\text{S}_2$ (M + H) 563.2714, found 563.2732.

Oleic Acid Benzyl Ester (7a) from Dimesylate 7. (a) Use of Li_2Te . Apart from a change in solvent, the general procedure was followed, using Te powder (200 mesh, 53 mg, 0.412 mmol), Et_3BHLi (1 M in THF, 0.78 mL, 0.78 mmol), dimesylate **7** (108 mg, 0.192 mmol) in dioxane (5 mL), and a reaction time of 14 h. Starting material was still present (TLC, silica, 40:60 CH_2Cl_2 –hexane), and so the mixture was heated at 100 $^\circ\text{C}$ for 2 h (TLC control). The mixture was cooled, washed out of the flask with hexane, and evaporated at room temperature. Flash chromatography of the residue over silica gel (1.5 \times 20 cm), using 2:3 CH_2Cl_2 –hexane, gave **7a** (59 mg, 83%) as a colorless oil, spectroscopically identical to an authentic sample.

(b) Use of Li_2Se . Se powder (325 mesh, 29 mg, 0.360 mmol) and a small stirring bar were placed in a dry round-bottomed flask fused onto a reflux condenser. The flask was closed with a septum and flushed with Ar. A solution of Et_3BHLi (1 M in THF, 0.68 mL, 0.68 mmol) was injected, and the mixture was stirred until a milky white suspension had formed (ca. 20 min). Dimesylate **7** (101 mg, 0.179 mmol) in dioxane (5 mL) was then injected dropwise, and the mixture was stirred for 24 h. Starting material was still present (TLC, silica, 40:60 CH_2Cl_2 –hexane), and so the mixture was heated at 100 $^\circ\text{C}$ for 4 h. The mixture was cooled, washed out of the flask with hexane, and evaporated at room temperature. Flash chromatography of the residue over silica gel (1 \times 30 cm), using 7:13 CH_2Cl_2 –hexane, gave **7a** (50.8 mg, 76%) as a colorless oil, spectroscopically identical to an authentic sample.

Methyl 5-O-Benzyl-2,3-di-O-mesyl- β -D-ribofuranoside (8). MeSO_2Cl (1.6 mL, 20 mmol) in CH_2Cl_2 (10 mL) was added dropwise to a stirred and cooled (0 $^\circ\text{C}$) solution of methyl 5-O-benzyl- β -D-ribofuranoside⁴⁴ (1.302 g, 5.122 mmol) and pyridine (3.3 mL, 41 mmol) in CH_2Cl_2 (10 mL). The ice bath was removed, and stirring was continued for 24 h. The mixture was poured onto ice (ca. 50 g) and extracted with EtOAc (100 mL). The organic extract was washed with aqueous CuSO_4 (10% w/v, 2 \times 50 mL), dried (MgSO_4), and evaporated. Flash chromatography of the residue over silica gel (3 \times 30 cm), using 3:7 EtOAc –hexane, gave **8** (1.963 g, 93%) as a colorless oil: FTIR (CH_2Cl_2 , cast) 1384, 1180 cm^{-1} ; ^1H NMR (CDCl_3 , 300 MHz) δ 2.97 (s, 3 H), 3.15 (s, 3 H), 3.41 (s, 3 H), 3.62 (dd, $J = 10.5$, 5.0 Hz, 1 H), 3.70 (dd, $J = 10.5$, 4.4 Hz, 1 H), 4.40 (ddd, $J = 6.5$, 5.0, 4.5 Hz, 1 H), 4.58 (q, $J = 17.0$, 12.0 Hz, 2 H), 4.98 (dd, $J = 5.0$, 1.5 Hz, 1 H), 5.08 (d, $J = 1.5$ Hz, 1 H), 5.20 (dd, $J = 6.5$, 5.0 Hz, 1 H), 7.28–7.45 (m, 5 H); ^{13}C NMR (CDCl_3 , 75.5 MHz) δ 38.12, 38.50, 55.65, 69.79, 73.74, 77.26, 79.15, 79.79, 105.68, 127.91, 128.02, 128.54, 137.49; exact mass m/z calcd for $\text{C}_{15}\text{H}_{22}\text{O}_9\text{S}_2$ 410.0706, found 410.0734.

Methyl 5-O-Benzyl-2,3-dideoxy- β -D-pent-2-enofuranoside (8a) from Dimesylate 8. The general procedure was followed, but at a different temperature for the second stage, using Te powder (200 mesh, 72 mg, 0.565 mmol), Et_3BHLi (1 M in THF, 1.27 mL, 1.27 mmol), and methyl 5-O-benzyl-2,3-di-O-mesyl- β -D-ribofuranoside (**8**) (100 mg, 0.244 mmol) in dioxane (5 mL). The mixture was refluxed for 20 h. At this stage all of the dimesylate had reacted (TLC, silica, 30:70 EtOAc –hexane). The mixture was cooled, washed out of the

(44) Using literature procedures, methyl 2,3-O-isopropylidene- β -D-ribofuranoside (Levene, P. A.; Stiller, E. T. *J. Biol. Chem.* **1934**, *104*, 299. Barrett, A. G. M.; Lebold, S. A. *J. Org. Chem.* **1990**, *55*, 3853) was converted into its 5-O-benzyl derivative (Tener, G. M.; Khorana, H. G. *J. Am. Chem. Soc.* **1957**, *79*, 437) and then into methyl 5-O-benzyl- β -D-ribofuranoside (Kawana, M.; Kuzuhara, H.; Emoto, S. *Bull. Chem. Soc. Jpn.* **1981**, *54*, 1492).

(43) Shonle, H. A.; Row, P. Q. *J. Am. Chem. Soc.* **1921**, *43*, 361.

flask with acetone, and evaporated at room temperature. Flash chromatography of the residue over silica gel (1 × 20 cm), using 1:9 EtOAc–hexane, gave **8a** (37 mg, 69%) as a colorless oil: FTIR (neat film) unexceptional; ¹H NMR identical to that reported;⁴⁵ ¹³C NMR (acetone-*d*₆, 75.5 MHz) δ 54.25, 73.54, 74.38, 85.31, 110.06, 128.11, 128.24, 128.33, 128.97, 133.74, 139.66; mass (CI) *m/z* calcd for C₁₃H₁₆O₃ 220, found 220.

5'-O-(Triphenylmethyl)uridine 2',3'-Bis(methanesulfonate) (9). (a) **5'-O-(Triphenylmethyl)uridine.** The literature procedure⁴⁶ was followed, but using a different workup.

Uridine (155 mg, 0.634 mmol), trityl chloride (199 mg, 0.715 mmol), and a small stirring bar were placed in a dry round-bottomed flask fused onto a reflux condenser. The flask was closed with a septum and flushed with Ar. Pyridine (1.90 mL) was injected, and the mixture was stirred at room temperature for 48 h. The mixture was then heated for 30 min (oil bath at 100 °C), cooled, and poured onto ice (*ca.* 25 g). The gummy product was filtered off, washed with water, and dissolved in acetone. Evaporation of the solvent and flash chromatography of the residue over silica gel (2 × 30 cm), using 1:19 MeOH–CH₂Cl₂, gave 5'-*O*-(triphenylmethyl)uridine (250 mg, 81%): mp 199–200 °C (lit.⁴⁶ mp 198–201 °C); FTIR (CH₂Cl₂, cast) 3600–3200, 1691 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz) δ 3.35–3.62 (m, 3 H), 4.10–4.25 (m, 1 H), 4.30–4.52 (m, 2 H), 5.25–5.38 (m, 1 H), 5.48–5.65 (br s, 1 H), 5.85–5.93 (m, 1 H), 7.18–7.50 (m, 15 H), 7.95–8.06 (apparent d, *J* = 8.0 Hz, 1 H), 10.35–10.45 (br s, 1 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 62.01, 69.71, 75.59, 83.80, 87.67, 90.88, 102.24, 127.48, 127.77, 128.09, 140.38, 143.24, 151.09, 163.77 (several signals of the trityl group overlap); exact mass *m/z* calcd for C₁₉H₁₅ (M – C₉H₁₁N₂O₆) 243.1174, found 243.1171; exact mass *m/z* calcd for C₉H₁₁N₂O₆ (M – C₁₀H₁₃) 243.0617, found 243.0620; mass (FAB) *m/z* calcd for C₂₈H₂₇N₂O₆ (M + H) 487, found 487.

(b) **5'-O-(Triphenylmethyl)uridine 2',3'-Bis(methanesulfonate) (9).** MeSO₂Cl (3.78 mL, 48.9 mmol) in dry CH₂Cl₂ (15 mL) was added dropwise to a stirred and cooled (0 °C) solution of 5'-*O*-(triphenylmethyl)uridine (5.944 g, 12.22 mmol) and dry pyridine (7.9 mL, 98 mmol) in dry CH₂Cl₂ (25 mL). The ice bath was removed, and stirring was continued for 15 h. The mixture was poured onto ice (*ca.* 200 g) and extracted with EtOAc (2 × 200 mL). The organic extract was washed with water (2 × 100 mL), aqueous NaOH (0.5 M, 2 × 100 mL), and aqueous CuSO₄ (10% w/v, 1 × 200 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue twice over silica gel (5 × 35 cm), using 1:49 MeOH–CH₂Cl₂ for the first chromatography and 1:99 MeOH–CH₂Cl₂ for the second, gave **9**⁴⁷ (5.837 g, 74%) as a colorless oil.

Another experiment, run on a smaller scale [5'-*O*-(triphenylmethyl)uridine, 2.282 g] gave the product in 86% yield: FTIR (CH₂Cl₂, cast) 1694, 1364, 1179 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz) δ 3.10 (s, 3 H), 3.21 (s, 3 H), 3.50–3.75 (m, 2 H), 4.25–4.50 (m, 1 H), 5.25–5.65 (m, 3 H), 6.02 (d, *J* = 3.0 Hz, 1 H), 7.10–7.60 (m, 15 H), 7.72 (d, *J* = 8.0 Hz, 1 H), 9.32 (br s, 1 H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 38.62, 38.86, 60.87, 73.58, 78.24, 80.93, 88.18, 88.43, 103.20, 127.69, 128.24, 128.73, 139.95, 142.68, 150.49, 163.02 (several signals of the trityl group overlap); mass (HRFAB) *m/z* calcd for C₃₀H₃₁N₂O₁₀S₂ (M + H) 643.1412, found 643.1401.

2',3'-Didehydro-2',3'-dideoxy-5'-O-(triphenylmethyl)uridine (9a) from Dimesylate 9. (a) **Use of Na₂Te.** Na₂Te (1.796 g, 10.35 mmol) (prepared from the elements, as described above) and a small stirring bar were placed in a dry 100-mL round-bottomed flask closed with a septum. The flask was flushed with Ar. Dimesylate **9** (2.660 g, 4.139 mmol) in dry THF (40 mL) was then injected, and the mixture was stirred for 20 h at room temperature. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temper-

ature. Flash chromatography of the residue over silica gel (4 × 30 cm), using 11:5:4 CH₂Cl₂–PhMe–MeCN, gave **9a** (1.402 g, 75%) as a colorless oil, spectroscopically identical to an authentic^{9b} sample.

Another experiment, run on a smaller scale (dimesylate, 139 mg), gave the product in 93% yield.

(b) **Use of Li₂Te in Dioxane–THF.** Apart from a change in solvent, the general procedure was followed, using Te powder (200 mesh, 40 mg, 0.313 mmol), Et₃BHLi (1 M in THF, 0.66 mL, 0.66 mmol), dimesylate **9** (100 mg, 0.155 mmol) in dioxane (5 mL), and a reaction time of 48 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (2 × 25 cm), using 10:7:3 CH₂Cl₂–PhMe–MeCN, gave **9a** (56 mg, 80%) as a colorless oil, spectroscopically identical to an authentic sample.

(c) **Use of Na₂Se.** Na₂Se (30 mg, 0.241 mmol) (prepared from the elements, as described above) and a small stirring bar were placed in a dry round-bottomed flask fused onto a reflux condenser. The flask was closed with a septum and flushed with Ar. Dimesylate **9** (59 mg, 0.092 mmol) in THF (2 mL) was then injected, and the mixture was stirred for 48 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue twice over silica gel (1 × 25 cm), using 11:5:4 CH₂Cl₂–PhMe–MeCN, gave **9a** (21.3 mg, 51%) as a colorless oil, spectroscopically identical to an authentic sample.

(d) **Use of Li₂Se.** Se powder (325 mesh, 15 mg, 0.187 mmol) and a small stirring bar were placed in a dry round-bottomed flask fused onto a reflux condenser. The flask was closed with a septum and flushed with Ar. A solution of Et₃BHLi (1 M in THF, 0.37 mL, 0.37 mmol) was injected, and the mixture was stirred for *ca.* 4 h. A milky white suspension was formed after 10 min. Dimesylate **9a** (59 mg, 0.093 mmol) in THF (3 mL) was then injected dropwise, and the mixture was stirred for 20 h. The mixture turned brown on initial addition of the dimesylate solution. The mixture was washed out of the flask with CH₂Cl₂, K₂CO₃ (500 mg) was added, and the mixture was then evaporated at room temperature. Flash chromatography of the residue over silica gel (1 × 20 cm), using 10:7:3 CH₂Cl₂–PhMe–MeCN, gave **9a** (27 mg, 65%) as a colorless oil, spectroscopically identical to an authentic sample.

(e) **Use of Li₂Te in the Presence of Acetonitrile.** Apart from a change in solvent, the general procedure was followed, using Te powder (200 mesh, 42 mg, 0.327 mmol), Et₃BHLi (1 M in THF, 0.73 mL, 0.73 mmol), dimesylate **9** (100 mg, 0.155 mmol) in dry MeCN (2 mL), and a reaction time of 16 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (1 × 30 cm), using 11:5:4 CH₂Cl₂–PhMe–MeCN, gave **9a** (70 mg, 99%) as a colorless oil, spectroscopically identical to an authentic sample.

(f) **Use of Na₂Te in the Presence of MeCN.** Dimesylate **9** (1.287 g, 2.003 mmol) in MeCN (40 mL) was added to Na₂Te (869 mg, 5.01 mmol, weighed out in a glove bag), and the mixture was stirred at room temperature for 24 h (Ar atmosphere). Evaporation of the solvent and flash chromatography of the residue over silica gel (4 × 20 cm), using 11:5:4 CH₂Cl₂–PhMe–MeCN, gave **9a** (400 mg, 44%) as a colorless oil, spectroscopically identical to an authentic sample.

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(49) Fox, J. J.; Yung, N.; Bendich, A. *J. Am. Chem. Soc.* **1957**, *79*, 2775. We prepared the compound (59% yield) by the literature procedure but ran the reaction at room temperature (24 h). The compound had: FTIR (CH₂Cl₂, cast) 3600–3200, 1691 cm⁻¹; ¹H NMR (acetone-*d*₆, 200 MHz) δ 1.48 (apparent s, 3 H), 3.35–3.48 (m, 2 H), 4.08–4.18 (m, 1 H), 4.32–4.48 (m, 3 H), 4.68–4.77 (m, 1 H), 5.90–6.02 (d, *J* = 4.0 Hz, 1 H), 7.22–7.62 (m, 16 H), 10.02 (br s, 1 H); ¹³C NMR (pyridine-*d*₅, 100.6 MHz) δ 12.41, 64.55, 71.50, 75.36, 84.10, 87.51, 90.06, 110.72, 127.61*, 128.38*, 129.17*, 136.05, 144.37*, 152.13, 164.83 (the four starred signals represent 18 carbons in trityl group); exact mass *m/z* calcd for C₂₉H₂₆N₂O₅ (M – H₂O) 482.1842, found 482.1843.

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5-Methyl-5'-O-(triphenylmethyl)uridine 2',3'-Bis(methanesulfonate) (10). The following procedure differs from the reported one.⁴⁸

MeSO₂Cl (0.11 mL, 1.4 mmol) in CH₂Cl₂ (1 mL) was added dropwise to a stirred and cooled (0 °C) solution of 5-methyl-5'-O-(triphenylmethyl)uridine⁴⁹ (174 mg, 0.348 mmol) and pyridine (0.46 mL, 5.6 mmol) in CH₂Cl₂ (3 mL). The ice bath was removed, and stirring was continued for 48 h. The mixture was poured onto ice (ca. 50 g) and extracted with EtOAc (2 × 50 mL). The organic extract was washed with water (2 × 50 mL), aqueous NaOH (0.1 M, 1 × 50 mL), and aqueous CuSO₄ (10% w/v), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (2 × 30 cm), using 3:97 MeOH-CH₂Cl₂, gave **10** (190 mg, 83%); mp 108–112 °C (lit.⁴⁸ mp 107–110 °C); FTIR (CH₂Cl₂, cast) 1693, 1364, 1180 cm⁻¹; ¹H NMR identical to that reported;⁴⁸ ¹³C NMR (CDCl₃, 50.3 MHz) δ 11.75, 38.68, 38.84, 61.92, 74.96, 77.24, 81.48, 87.83, 88.23, 112.38, 127.71, 128.22, 128.77, 135.31, 142.92, 150.64, 163.39 (several signals of the trityl group overlap); mass (HRFAB) *m/z* calcd for C₃₁H₃₃N₂O₁₀S₂ (M + H) 657.1578, found 657.1548.

2',3'-Didehydro-2',3'-dideoxy-5-methyl-5'-O-(triphenylmethyl)uridine (10a) from Dimesylate 10. The general procedure was followed, using Te powder (200 mesh, 25 mg, 0.195 mmol), Et₃BHLi (1 M in THF, 0.50 mL, 0.50 mmol), dimesylate **10** (60 mg, 0.092 mmol) in THF (3 mL), and a reaction time of 48 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (2 × 30 cm), using 10:7:3 CH₂Cl₂-PhMe-MeCN, gave **10a** (38.6 mg, 90%); FTIR (CH₂Cl₂, cast) 1689 cm⁻¹; ¹H NMR identical to that reported;^{35b} ¹³C NMR (CDCl₃, 100.6 MHz) δ 11.40, 64.96, 85.72, 87.03, 89.71, 111.28, 126.35, 127.41, 127.97, 128.79, 134.56, 136.03, 143.25, 150.80, 163.82 (several signals of the trityl group overlap); exact mass *m/z* calcd for C₂₀H₁₇O (M - C₉H₉N₂O₃) 273.1279, found 273.1281; exact mass *m/z* calcd for C₉H₉N₂O₃ (M - C₂₀H₁₇O) 193.0613, found 193.0611.

5'-O-(Triphenylmethyl)uridine 2',3'-Bis(*p*-toluenesulfonate) (11). *p*-TsCl (470 mg, 1.47 mmol) in CH₂Cl₂ (2 mL) was added dropwise to a stirred and cooled (0 °C) solution of 5'-O-(triphenylmethyl)uridine⁴⁶ (200 mg, 0.411 mmol), pyridine (0.80 mL, 9.9 mmol), and 4-(dimethylamino)pyridine (5 mg) in CH₂Cl₂ (2 mL). The ice bath was removed, stirring was continued for 24 h, and the mixture was then heated at 50 °C for a further 24 h. The mixture was cooled, poured onto ice (ca. 25 g), and extracted with CH₂Cl₂ (1 × 100 mL). The organic extract was washed with aqueous CuSO₄ (10% w/v, 2 × 50 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (1 × 30 cm), using 11:5:4 CH₂Cl₂-PhMe-MeCN, gave **11** (93 mg, 29%) as a colorless oil, which was used in the next step without full characterization: ¹H NMR (CDCl₃, 200 MHz) δ 2.40 (s, 3 H), 2.45 (s, 3 H), 3.30–3.50 (m, 2 H), 4.30–4.45 (m, 1 H), 5.00–5.35 (m, 3 H), 6.10 (d, *J* = 6.0 Hz, 1 H), 7.15–7.45 (m, 20 H), 7.64 (d, *J* = 8.0 Hz, 2 H), 7.77 (d, *J* = 8.0 Hz, 2 H), 7.96 (br s, 1 H).

2',3'-Didehydro-2',3'-dideoxy-5'-O-(triphenylmethyl)uridine (9a) from Bis(*p*-toluenesulfonate) 11. The general procedure was followed, using Te powder (200 mesh, 12 mg, 0.092 mmol), Et₃BHLi (1 M in THF, 0.21 mL, 0.21 mmol), ditosylate **11** (34.8 mg, 0.0438 mmol) in THF (1 mL), and a reaction time of 24 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (1 × 30 cm), using 10:5:4 CH₂Cl₂-PhMe-MeCN, gave **9a** (11.8 mg, 60%) as a colorless oil, spectroscopically identical to an authentic sample.

***N*-Acetyl-5'-O-(triphenylmethyl)cytidine.** A very similar procedure (reaction temperature 110 °C) has been reported,⁵⁰ but the yield was only 14%.

Pyridine (10 mL) was injected onto a stirred mixture of *N*-acetylcytidine⁵¹ (1.000 g, 3.506 mmol) and trityl chloride (1.075 g, 3.856 mmol), and stirring was continued for 36 h (Ar atmosphere). The mixture was evaporated, diluted with CH₂

Cl₂, and again evaporated. The gummy residue was washed with water, and the residue was dissolved in acetone. The solution was dried (MgSO₄) and evaporated. Flash chromatography of the residue over silica gel (4.5 × 30 cm), using 7:93 MeOH-CH₂Cl₂, gave *N*-acetyl-5'-O-(triphenylmethyl)cytidine (1.475 g, 80%) as a colorless oil: FTIR (CH₂Cl₂, cast) 3600–3200, 1656 cm⁻¹; ¹H NMR (CDCl₃, 200 MHz) δ 2.25 (s, 3 H), 3.30–3.55 (m, 3 H), 4.30–4.48 (m, 3 H), 5.58 (br s, 1 H), 5.80–5.90 (m, 1 H), 7.20–7.40 (m, 16 H), 8.10–8.20 (d, *J* = 8.0 Hz, 1 H), 8.55–8.77 (br s, 1 H); ¹³C NMR (CDCl₃, 100.6 MHz) δ 24.81, 62.12, 69.88, 76.02, 84.04, 87.44, 92.24, 96.98, 127.32, 127.98, 128.57, 143.18, 144.73, 156.23, 162.64, 170.74 (several signals of the trityl group overlap); exact mass *m/z* calcd for C₃₀H₂₇N₃O₅ (M - H₂O) 509.1951, found 509.1943.

***N*-Acetyl-5'-O-(triphenylmethyl)cytidine 2',3'-Bis(methanesulfonate) (12).** MeSO₂Cl (0.061 mL, 0.786 mmol) in CH₂Cl₂ (0.6 mL) was added dropwise to a stirred and cooled (0 °C) solution of *N*-acetyl-5'-O-(triphenylmethyl)cytidine (104 mg, 0.196 mmol) and Et₃N (0.060 mL, 0.434 mmol) in CH₂Cl₂ (1 mL). The mixture was stirred at 0 °C for 25 min, poured onto ice (ca. 100 g), and extracted with CH₂Cl₂ (100 mL). The organic extract was washed with water (1 × 100 mL), saturated aqueous NaHCO₃ (1 × 100 mL), and water (1 × 100 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (1 × 30 cm), using 3.5:96.5 MeOH-CH₂Cl₂, gave **12** (108 mg, 81%); FTIR (CH₂Cl₂, cast) 1722, 1666, 1490, 1366, 1181 cm⁻¹; ¹H NMR (CD₂Cl₂, 200 MHz) δ 2.18 (s, 3 H), 3.06 (s, 3 H), 3.35 (s, 3 H), 3.56 (dd, *J* = 11.5, 2.2 Hz, 1 H), 3.68 (dd, *J* = 11.5, 2.2 Hz, 1 H), 4.30–4.50 (m, 1 H), 5.37–5.55 (m, 2 H), 5.97 (br s, 1 H), 7.10 (d, *J* = 7.0 Hz, 1 H), 7.20–7.60 (m, 15 H), 8.25 (d, *J* = 7.0 Hz, 1 H), 8.90 (br s, 1 H); ¹³C NMR (CD₂Cl₂, 100.6 MHz) δ 25.01, 38.90, 39.24, 60.49, 72.58, 79.89, 80.58, 88.22, 90.44, 97.46, 127.87, 128.47, 129.02, 143.30, 144.85, 155.29, 163.71, 171.23 (several signals of the trityl group overlap); mass (HRFAB) *m/z* calcd for C₃₂H₃₄N₃O₁₀S₂ (M + H) 684.1687, found 684.1651.

***N*-Acetyl-2',3'-didehydro-2',3'-dideoxy-5'-O-(triphenylmethyl)cytidine (12a) from Dimesylate 12.** (a) **Use of Li₂Te.** The general procedure was followed, using Te powder (200 mesh, 39 mg, 0.307 mmol), Et₃BHLi (1 M in THF, 0.66 mL, 0.66 mmol), *N*-acetyl-5'-O-(triphenylmethyl)cytidine 2',3'-dimethanesulfonate (100 mg, 0.146 mmol) in THF (2 mL), and a reaction time of 14 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (1 × 15 cm), using 5:3:2 MeCN-CH₂Cl₂-PhMe, gave **12a** (60 mg, 83%) as a colorless oil containing trace impurities (¹H NMR, 200 MHz): FTIR (CH₂Cl₂, cast) 1722, 1672 cm⁻¹; ¹H NMR (CD₂Cl₂, 200 MHz) δ 2.18 (s, 3 H), 3.30–3.50 (m, 2 H), 4.95–5.15 (m, 1 H), 5.93–6.08 (m, 1 H), 6.18–6.43 (m, 1 H), 6.87 (d, *J* = 7.0 Hz, 1 H), 6.92–7.05 (m, 1 H), 7.15–7.55 (m, 15 H), 8.0 (d, *J* = 7.0 Hz, 1 H), 8.92 (br s, 1 H); ¹³C NMR (CD₂Cl₂, 100.6 MHz) δ 25.04, 65.21, 86.84, 87.61, 92.00, 96.77, 127.42, 127.68, 128.34, 129.04, 133.71, 143.78, 145.70, 155.80, 163.30, 171.03 (several signals of the trityl group overlap); exact mass *m/z* calcd for C₁₉H₁₅ (M - C₁₁H₁₂N₃O₄) 243.1174, found 243.1173; exact mass *m/z* calcd for C₁₁H₁₂N₃O₄ (M - C₁₉H₁₅) 250.0828, found 250.0836.

(b) **Use of Na₂Te.** Na₂Te (680 mg, 0.393 mmol) (prepared from the elements, as described above) and a small stirring bar were placed in a dry round-bottomed flask fused onto a reflux condenser. The flask was closed with a septum and flushed with Ar. *N*-Acetyl-5'-O-(triphenylmethyl)cytidine 2',3'-bis(methanesulfonate) (107 mg, 0.157 mmol) in THF (2 mL) was then injected, and the mixture was stirred for 24 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (1 × 25 cm), using 5:3:2 MeCN-CH₂Cl₂-PhMe, gave **12a** (33 mg, 42%) as a colorless oil, spectroscopically identical to an authentic sample.

***N*-Acetyl-2',3'-dideoxy-5'-O-(triphenylmethyl)cytidine by Hydrogenation of 12a.** *N*-Acetyl-2',3'-didehydro-2',3'-dideoxy-5'-O-(triphenylmethyl)cytidine (**12a**) (50 mg, 0.101 mmol), EtOAc (3 mL), and MeOH (1 mL) were placed in a test tube together with Pd/charcoal (10% w/w, 10 mg). The test tube was supported with glass wool in a Parr vessel and

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shaken with hydrogen (50 psi) for 4 h. The mixture was filtered and evaporated. Flash chromatography of the residue over silica gel (1 × 30 cm), using 5:3:2 MeCN-CH₂Cl₂-PhMe, gave *N*-acetyl-2',3'-dideoxy-5'-*O*-(triphenylmethyl)cytidine (30 mg, 61%) as a colorless oil: FTIR (CH₂Cl₂, cast) 3229, 2925, 1717, 1663, 1493 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ 1.88–1.98 (m, 2 H), 2.09–2.20 (m, 4 H), 2.41–2.55 (m, 1 H), 3.36 (dd, *J* = 4.5, 4.5 Hz, 1 H), 3.49 (dd, *J* = 2.7, 2.7 Hz, 1 H), 4.22–4.32 (m, 1 H), 6.03 (dd, *J* = 2.0, 2.0 Hz, 1 H), 7.10 (d, *J* = 7.5 Hz, 1 H), 7.25–7.49 (m, 15 H), 8.31 (d, *J* = 7.5 Hz, 1 H), 9.12 (br s, 1 H); ¹³C NMR (CD₂Cl₂, 75 MHz) δ 25.08 (q'), 25.17 (t'), 38.87 (t'), 64.38 (t'), 82.20 (d'), 87.63 (s'), 88.32 (d'), 95.48 (d'), 127.67 (d'), 128.38 (d'), 129.05 (d'), 144.02 (s'), 145.31 (d'), 155.36 (s'), 162.71 (s'), 170.50 (s') (several signals of the trityl group overlap); mass (FAB) *m/z* calcd for C₃₀H₃₀N₃O₄ (M + H) 496.2, found 496.1.

5'-*O*-Acetyluridine 2',3'-Bis(methanesulfonate) (13). MeSO₂Cl (0.90 mL, 11.60 mmol) in CH₂Cl₂ (1.6 mL) was added dropwise to a stirred and cooled (ice bath) solution of 5'-*O*-acetyluridine^{52,53} (332 mg, 1.16 mmol) and pyridine (1.50 mL, 18.5 mmol) in CH₂Cl₂ (3 mL). The ice bath was removed, and stirring was continued for 24 h. The mixture was evaporated at room temperature, and flash chromatography of the residue over silica gel (3.5 × 30 cm), using 3:97 MeOH-CH₂Cl₂, gave, after a second chromatography under the same conditions, **13** (417 mg, 81%); FTIR (MeOH, cast) 1365, 1180 cm⁻¹; ¹H NMR (acetone-*d*₆, 200 MHz) δ 2.07 (s, 3 H), 3.26 (s, 3 H), 3.32 (s, 3 H), 4.30–4.60 (m, 3 H), 5.48 (t, *J* = 6.0 Hz, 1 H), 5.55–5.80 (m, 2 H), 6.00 (d, *J* = 3.0 Hz, 1 H), 7.76 (d, *J* = 8.0 Hz, 1 H), 10.23 (br s, 1 H); ¹³C NMR (acetone-*d*₆, 50.3 MHz) δ 20.61, 38.66, 38.74, 62.58, 75.14, 78.61, 80.24, 91.11, 103.34, 141.93, 151.33, 163.36, 170.57; mass (HRFAB) *m/z* calcd for (C₁₃H₁₈N₂O₁₁S₂ + H) 443.0431, found 443.0398.

5'-*O*-Acetyl-2',3'-dideoxy-2',3'-dideoxyuridine (13a) from Dimesylate 13. The general procedure was followed, using Te powder (200 mesh, 61 mg, 0.475 mmol), Et₃BHLi (1 M in THF, 1.17 mL, 1.17 mmol), dimesylate **13** (100 mg, 0.226 mmol) in THF (3 mL), and a reaction time of 96 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (1 × 30 cm), using 3:97 MeOH-CH₂Cl₂, gave **13a** (7.7 mg, 14%), spectroscopically identical to an authentic^{9b,54} sample.

5'-*O*-[(1,1-Dimethylethyl)dimethylsilyl]adenosine 2',3'-Bis(methanesulfonate) (14). MeSO₂Cl (0.57 mL, 7.3 mmol) in CH₂Cl₂ (2 mL) was added dropwise to a stirred and cooled (0 °C) solution of 5'-*O*-(1,1-dimethylethyl)dimethylsilyl[adenosine⁵⁵ (700 mg, 1.84 mmol) in a mixture of CH₂Cl₂ (2 mL) and pyridine (8.3 mL). After 1 h, the ice bath was removed, and stirring was continued for 24 h; the mixture was then poured onto ice, EtOAc (200 mL) was added, and the organic layer was dried (MgSO₄) and evaporated. Flash chromatography of the residue over silica gel (3 × 20 cm), using 1:19 MeOH-CH₂Cl₂, gave **14** (773 mg, 78%) as a colorless oil: FTIR (CH₂Cl₂, cast) 3315, 3148 cm⁻¹; ¹H NMR (acetone-*d*₆, 200 MHz) δ 0.07 (s, 3 H), 0.08 (s, 3 H), 0.88 (s, 9 H), 3.25 (s, 3 H), 3.32 (s, 3 H), 3.92–4.02 (dd, *J* = 12.0, 4.0 Hz, 1 H), 4.02–4.15 (dd,

J = 12.0, 4.0 Hz, 1 H), 4.38–4.48 (m, 1 H), 5.70–5.80 (apparent t, *J* = 5.0 Hz, 1 H), 6.08–6.18 (apparent t, *J* = 5.0 Hz, 1 H), 6.32–6.38 (d, *J* = 5.0 Hz, 1 H), 6.60–6.76 (br s, 2 H), 8.22 (s, 1 H), 8.25 (s, 1 H); ¹³C NMR (acetone-*d*₆, 50.3 MHz) δ -5.40 (two overlapping signals), 18.94, 26.28 (three overlapping signals), 38.67 (two overlapping signals), 62.29, 76.20, 78.31, 83.50, 87.15, 120.89, 140.49, 150.50, 153.96, 157.36; mass (FAB) *m/z* calcd for C₁₈H₃₂N₅O₈S₂Si (M + H) 538, found 538.

5'-*O*-[(1,1-Dimethylethyl)dimethylsilyl]-2',3'-dideoxy-2',3'-dideoxyadenosine (14a) from Dimesylate 14. The general procedure was followed, using Te powder (200 mesh, 113 mg, 0.887 mmol), Et₃BHLi (1 M in THF, 2.0 mL, 2.0 mmol), an initial reaction time of ca. 6 h, dimesylate **14** (200 mg, 0.372 mmol) in THF (2 mL), and a reaction time of 20 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (2 × 15 cm), using 1:19 MeOH-CH₂Cl₂, gave **14a**⁵⁶ (101 mg, 78%) as a colorless oil: FTIR (CH₂Cl₂, cast) 3292, 3164, 2927, 1601, 1085, 837 cm⁻¹; ¹H NMR (CD₂Cl₂, 400 MHz) δ 0.04 (s, 3 H), 0.05 (s, 3 H), 0.89 (s, 9 H), 3.82 (d, *J* = 4.0 Hz, 2 H), 4.94–5.00 (m, 1 H), 5.91 (br s, W_{1/2} = 17 Hz, 2 H), 6.04 (ddd, *J* = 6.0, 4.0, 2.0 Hz, 1 H), 6.41 (dt *J* = 6.0, 2.0, 2.0 Hz, 1 H), 7.04–7.08 (m, *J* = 2.0, 1 H), 8.08 (s, 1 H), 8.30 (s, 1 H); ¹³C (CD₂Cl₂, 75 MHz) δ -5.17 (q'), -5.06 (q') (two overlapping signals), 18.95 (s'), 26.29 (q') (three overlapping signals), 65.45 (t'), 88.27 (d'), 88.68 (d'), 120.19 (s'), 126.06 (d'), 134.96 (d'), 139.81 (d'), 150.44 (s'), 153.58 (d'), 156.01 (s'); exact mass *m/z* calcd for C₁₆H₂₅N₅O₂Si 347.1778, found 347.1776.

***N*-[(Dimethylamino)methylene]-5'-*O*-[bis(4-methoxyphenyl)phenylmethyl]adenosine 2',3'-Bis(methanesulfonate) (15).** MeSO₂Cl (0.37 mL, 4.8 mmol) in CH₂Cl₂ (5 mL) was added dropwise to a stirred and cooled (0 °C) solution of *N*-[(dimethylamino)methylene]-5'-*O*-[bis(4-methoxyphenyl)phenylmethyl]adenosine,⁵⁷ (1.001 g, 1.602 mmol), and Et₃N (1.34 mL, 9.61 mmol) in CH₂Cl₂ (8 mL). The mixture was stirred at 0 °C for 30 min, poured onto ice (ca. 200 g), and extracted with CH₂Cl₂ (2 × 150 mL). The organic extract was washed with water (1 × 100 mL), saturated aqueous NaHCO₃ (1 × 100 mL), and water (1 × 100 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (3 × 30 cm), using 49:30:20:1 CH₂Cl₂-PhMe-MeCN-Et₃N, gave **15** (1.089 g, 87%); FTIR (CH₂Cl₂, cast) 1365, 1180 cm⁻¹; ¹H NMR (acetone-*d*₆, 200 MHz) δ 3.10–3.30 (m, 12 H), 3.40 (dd, *J* = 11.0, 4.0 Hz, 1 H), 3.62 (dd, *J* = 11.0, 3.5 Hz, 1 H), 3.75 (s, 3 H), 3.76 (s, 3 H), 4.40–4.60 (m, 1 H), 5.85–6.00 (m, 1 H), 6.30–6.55 (m, 2 H), 6.70–6.90 (m, 4 H), 7.10–7.38 (m, 7 H), 7.38–7.55 (m, 2 H), 8.30 (s, 1 H), 8.31 (s, 1 H), 8.85–9.00 (br s, 1 H); ¹³C NMR (acetone-*d*₆, 50.3 MHz) δ 35.02, 38.74, 38.80, 41.13, 55.60 (two overlapping signals), 62.99, 77.08, 77.47, 82.51, 87.67, 87.86, 114.06, 127.68, 128.63, 129.14, 131.04, 136.53, 136.65, 145.76, 152.45, 153.25, 159.14, 159.82, 161.16 (several signals of DMT group overlap); mass (HRFAB) *m/z* calcd for (C₃₆H₄₀N₆O₁₀S₂ + H) 781.2328, found 781.2337.

2',3'-Dideoxy-2',3'-dideoxy-*N*-[(dimethylamino)methylene]-5'-*O*-[bis(4-methoxyphenyl)phenylmethyl]adenosine (15a) from Dimesylate 15. The general procedure was followed, using Te powder (200 mesh, 20 mg, 0.156 mmol), Et₃BHLi (1 M in THF, 0.33 mL, 0.33 mmol), dimesylate **15** (58 mg, 0.074 mmol) in THF (2 mL), and a reaction time of 16 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated at room temperature. Flash chromatography of the residue over silica gel (1 × 30 cm), using 29:20:50:1 CH₂Cl₂-PhMe-MeCN-Et₃N, gave **15a** (39 mg, ca. 89%), containing trace impurities (¹H NMR, 200 MHz): ¹H NMR (acetone-*d*₆, 200 MHz) δ 3.02–3.30 (m, 6 H), 3.30–3.46 (m, 1 H), 3.46–3.65 (m, 1 H), 3.74 (s, 3 H), 3.75 (s, 3 H), 5.02–5.18 (m, 1 H), 6.15–6.30 (m, 1 H), 6.48–6.60 (m, 1 H), 6.60–6.90 (m, 4 H), 7.00–7.32 (m, 8 H), 7.32–7.50 (m, 2 H), 8.01 (s, 1 H), 8.42 (s, 1 H), 8.86–9.05 (s, 1 H). For characterization of the material was processed as described in the next experiment.

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(53) 2',3'-*O*-Isopropylideneuridine, prepared (91%) by the reported procedure (ref 52), except that the product was isolated by flash chromatography (silica gel, 1:19 MeOH-CH₂Cl₂), had: FTIR (CH₂Cl₂, cast) 3600–3300, 3300–3140, 1692 cm⁻¹; ¹H NMR (acetone-*d*₆, 200 MHz) δ 1.33 (s, 3 H), 1.53 (s, 3 H), 3.72–3.82 (apparent t, *J* = 7.6, 3.8 Hz, 2 H), 4.00–4.40 (m, including br s at δ 4.23, 2 H in all), 4.82–5.00 (m, 2 H), 5.55–5.65 (m, 1 H), 5.88–5.95 (d, *J* = 3.6 Hz, 1 H), 7.85–7.88 (d, *J* = 8.0 Hz, 1 H), 9.60–10.30 (br s, 1 H); ¹³C NMR (acetone-*d*₆, 50.3 MHz) δ 25.57, 27.52, 63.06, 67.97, 80.96, 84.49, 87.43, 95.58, 102.90, 142.96, 150.90, 160.13; exact mass *m/z* calcd for C₁₂H₁₆N₂O₆ 284.1008, found 284.1011. 5'-*O*-Acetyluridine, prepared from 2',3'-*O*-isopropylideneuridine, had: ¹H NMR (acetone-*d*₆, 200 MHz) δ 2.08 (s, 3 H), 4.10–4.38 (m, 5 H), 4.38–4.58 (br s, 1 H), 4.60–4.85 (br s, 1 H), 5.60–5.70 (d, *J* = 8.0 Hz, 1 H), 5.82–5.92 (d, *J* = 4.0 Hz, 1 H), 7.62–7.72 (d, *J* = 8.0 Hz, 1 H), 9.75–10.25 (br s, 1 H).

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5'-O-[Bis(4-methoxyphenyl)phenylmethyl]-2',3'-dideoxyadenosine and Use of β -Cyclodextrin for Tellurium Recovery. Te powder (200 mesh, 34 mg, 0.270 mmol) and a small stirring bar were placed in a dry round-bottomed flask fused onto a reflux condenser. The flask was closed with a septum and flushed with Ar. A solution of Et_3BHLi (1 M in THF, 0.60 mL, 0.60 mmol) was injected, and the mixture was stirred at 40 °C for 0.5 h, by which time the mixture had turned white. Dimesylate **15** (100 mg, 0.128 mmol) in THF (2 mL) was added to the resulting Li_2Te suspension. The mixture was stirred at room temperature for 14 h, and then β -cyclodextrin (500 mg) was added, together with 5:2.9:2:0.1 MeCN- CH_2Cl_2 -PhMe- Et_3N (20 mL). The resulting mixture was evaporated to deposit the Te on the β -cyclodextrin and then filtered through a pad of β -cyclodextrin (3 \times 3 cm), using the above solvent system. Flash chromatography silica gel (1 g) was added to the filtrate, and the mixture was evaporated. MeOH (20 mL) was added, and the mixture was again filtered, using MeOH. The filtrate was evaporated, and MeOH (4 mL) was added, together with Pd/carbon (20 mg). The mixture was placed under H_2 (50 psi) and shaken for 3 h, filtered, and evaporated. At this stage hydrogenation was incomplete (^1H NMR). MeOH (4 mL) and Pd/carbon (20 mg) were again added, and the reaction was set up as before and continued for 12 h, at which point hydrogenation was complete (^1H NMR). THF (4 mL) and concentrated NH_4OH (0.4 mL) were added, and the mixture was stirred for 1 h. Evaporation of the solvent and flash chromatography of the residue over silica gel (1 \times 15 cm), using 5:2.9:2:0.1 MeCN- CH_2Cl_2 -PhMe- Et_3N , gave 5'-O-[bis(4-methoxyphenyl)phenylmethyl]-2',3'-dideoxyadenosine⁵⁸ (30 mg, 43%) as a colorless oil: FTIR (CH_2Cl_2 , cast) 3320, 3180 cm^{-1} ; ^1H NMR (acetone- d_6 , 200 MHz) δ 2.10–2.35 (m, 2 H), 2.45–2.75 (m, 2 H), 3.22–3.38 (m, 2 H), 3.75 (s, 6 H), 4.28–4.44 (m, 1 H), 6.28–6.38 (m, 1 H), 6.65–6.88 (m, 6 H), 7.10–7.48 (m, 9 H), 8.15 (s, 1 H), 8.19 (s, 1 H); ^{13}C NMR (acetone- d_6 , 50.3 MHz) δ 27.54, 32.51, 55.52, 66.50, 81.44, 86.07, 86.44, 113.88, 127.48, 128.51, 129.07, 130.92, 137.00, 146.23, 150.46, 153.50, 157.06, 159.64 (several signals of DMT group overlap); exact mass m/z calcd for $\text{C}_{31}\text{H}_{31}\text{N}_5\text{O}_4$ 537.2376, found 537.2374.

5'-O-(4,4-Dimethoxytrityl)inosine 2',3'-Bis(methanesulfonate) (16). (a) **5'-O-(4,4-Dimethoxytrityl)inosine.** Pyridine (2 mL) was injected into a stirred solution of inosine (268.2 mg, 1.0 mmol) and 4,4'-dimethoxytrityl chloride (440.5 mg, 1.30 mmol) in dry DMSO (6 mL). Stirring was continued for 12 h. The mixture was poured onto ice and extracted with CHCl_3 . The combined organic extracts were washed successively with saturated aqueous NaHCO_3 (5 mL), water (2 \times 10 mL), and brine (5 mL). The organic solution was dried (MgSO_4), diluted with PhMe and evaporated (water pump). The residue was taken up in PhMe, and again evaporated. Finally, the residue was kept under oil pump vacuum to remove traces of PhMe. The crude 5'-O-protected inosine was used directly in the next step.

(b) **5'-O-(4,4-Dimethoxytrityl)inosine 2',3'-Bis(methanesulfonate) (16).** MeSO_2Cl (0.23 mL, 2.97 mmol) in CH_2Cl_2 (3.0 mL) was added dropwise over 20 min to a stirred and cooled (0 °C) solution of 5'-O-(4,4-dimethoxytrityl)inosine (432.1 mg, 0.7573 mmol) in pyridine (8 mL). After 10 min, the ice bath was removed, and stirring was continued for 3.5 h. The mixture was poured onto ice (ca. 10 g) and extracted with CH_2Cl_2 (100 mL). The organic extract was washed with water (10 mL), aqueous saturated NaHCO_3 (2 \times 5 mL), water (10 mL), and brine, dried (MgSO_4), diluted with PhMe, and evaporated (water pump). Evaporation from PhMe was repeated twice more. Finally, the residue was kept under oil pump vacuum to remove traces of PhMe. Flash chromatography of the residue over silica gel (2.5 \times 20 cm), using 90:8:2 CHCl_3 -MeOH-MeCN, gave **16** (402.2 mg, 59% over two steps) as a colorless oil: FTIR (CH_2Cl_2 , cast) 3057, 2934, 1699, 1510, 1367, 1251 cm^{-1} ; ^1H NMR (CD_2Cl_2 , 300 MHz) δ 3.07 (s, 3 H), 3.09 (s, 3 H), 3.41 (dd, J = 11.5, 3.8 Hz, 1 H), 3.61 (dd, J = 11.5, 3.0 Hz, 1 H), 3.75 (s, 3 H), 3.76 (s, 3 H), 4.48 (dd, J =

8.3, 3.5 Hz, 1 H); 5.61 (dd, J = 5.5, 5.0 Hz, 1 H), 6.04 (t, J = 5.5 Hz, 1 H), 6.23 (d, J = 5.5 Hz, 1 H); 6.75–7.50 (m, 13 H), 7.95 (s, 1 H), 8.02 (s, 1 H), 12.25 (s, 1 H); ^{13}C NMR (CD_2Cl_2 , 75 MHz) δ 38.72 (q'), 55.61 (q'), 62.40 (t'), 76.15 (d'), 77.17 (d'), 82.44 (d'), 86.74 (d'), 87.34 (s'), 113.61 (d'), 126.18 (s'), 127.46 (d'), 128.33 (d'), 128.43 (d'), 130.46 (d'), 135.45 (s'), 135.60 (s'), 144.75 (s'), 158.63 (s'), 159.24 (s') (the peaks corresponding to carbons 2, 4, and 8 in the purine base were not detected under the experimental conditions employed, and several signals of the DMT group overlap); exact mass m/z calcd for $\text{C}_{21}\text{H}_{19}\text{O}_2$ (M - $\text{C}_{12}\text{H}_{15}\text{O}_9\text{N}_4\text{S}_2$) 303.1385, found 303.1383.

(c) **5'-O-(4,4-Dimethoxytrityl)-2',3'-didehydro-2',3'-dideoxyinosine (16a) from Dimesylate 16.** The general procedure was followed, using Te powder (oven dried at 120 °C for 24 h, 200 mesh, 45.1 mg, 0.3534 mmol), Et_3BHLi (1 M in THF, 0.74 mL, 0.74 mmol), an initial reaction time of 1 h at 40 °C, 5'-O-(4,4-dimethoxytrityl)inosine 2',3'-bis(methanesulfonate) (100 mg, 0.138 mmol) in THF (5 mL). The mixture was stirred at 40 °C for 24 h. At this stage all of the dimesylate had reacted (TLC, silica, 7.5:1.8:0.7 CH_2Cl_2 -BuOAc-MeOH). The mixture was cooled, washed out of the flask with CH_2Cl_2 , and evaporated at room temperature. Flash chromatography of the residue over silica gel (2.5 \times 15 cm), using 3:47 MeOH- CH_2Cl_2 , gave **16a** (68.8 mg, ca. 93%) as an off-white solid containing slight impurities (^1H NMR, 300 MHz): FTIR (CH_2Cl_2 , cast) 3058, 2909, 1703, 1509, 1250 cm^{-1} ; ^1H NMR (CD_2Cl_2 , 300 MHz) δ 3.22 (dd, J = 4.0, 4.0 Hz, 1 H), 3.38 (dd, J = 6.3, 6.3 Hz, 1 H), 3.8 (s, 6 H), 5.1 (br s, 1 H), 6.09 (d, J = 5.8 Hz, 1 H), 6.42 (d, J = 6.0 Hz, 1 H), 6.70–6.80 (m, 4 H), 7.05 (br s, $W_{1/2}$ = 6.4 Hz, 1 H), 7.15–7.42 (m, 9 H), 7.85 (s, 1 H), 8.28 (s, 1 H), 12.84 (s, 1 H); ^{13}C NMR (CD_2Cl_2 , 100 MHz) δ 55.55 (q'), 65.92 (t'), 86.68 (s'), 87.05 (d'), 89.06 (d'), 113.04 (d'), 125.05 (s'), 125.32 (d'), 127.18 (d'), 128.14 (d'), 128.41 (d'), 130.33 (d'), 130.38 (d'), 135.14 (d'), 135.91 (s'), 136.00 (s'), 138.75 (d'), 145.08 (d'), 145.77 (d'), 149.33 (s'), 159.04 (s'), 159.29 (s') (several signals of DMT group overlap); mass (FAB) m/z calcd for $\text{C}_{31}\text{H}_{28}\text{N}_4\text{O}_5$ (M) 536.2, found 536.3.

5,6-Bis-O-(methylsulfonyl)-1,2-O-isopropylidene-3-O-methyl- α -D-glucufuranose (17). MeSO_2Cl (1.82 mL, 23.7 mmol) was added dropwise over 50 min to a stirred and cooled (ice bath) solution of 1,2-O-isopropylidene-3-O-methyl- α -D-glucufuranose⁵⁹ (1.11 g, 4.74 mmol) and pyridine (3.10 mL, 37.9 mmol) in CH_2Cl_2 . The mixture was stirred at 0 °C for 10 min, and the cold bath was then removed. Stirring was continued for 18 h, and the mixture was poured onto ice (ca. 50 g), and extracted with EtOAc (3 \times 35 mL). The combined organic extracts were washed with saturated aqueous NaHCO_3 (2 \times 20 mL), aqueous CuSO_4 (10% w/v, 2 \times 50 mL), and brine, dried (MgSO_4), and evaporated. Flash chromatography of the residue over silica gel (5 \times 15 cm), using 7:3 EtOAc-hexane, gave the dimesylate (1.45 g, 78%) as a crystalline solid: mp 78–79 °C; FTIR (CH_2Cl_2 , cast) 3000, 1350, 1180 cm^{-1} ; ^1H NMR (CDCl_3 , 200 MHz) δ 1.31 (s, 3 H), 1.49 (s, 3 H), 3.06 (s, 3 H), 3.12 (s, 3 H), 3.45 (s, 3 H), 3.85 (d, J = 1.8, 1 H), 4.36 (dd, J = 9.0, 3.0 Hz, 1 H), 4.44 (dd, J = 9.0, 5.4 Hz, 1 H), 4.60 (d, J = 3.0 Hz, 1 H), 4.66 (dd, J = 9.0, 1.9 Hz, 1 H), 5.16–5.21 (m, 1 H), 5.89 (d, J = 3.0 Hz, 1 H); ^{13}C NMR (CDCl_3 , 75.5 MHz) δ 26.26 (q'), 26.90 (q'), 37.70 (q'), 39.16 (q'), 57.79 (q'), 68.99 (t'), 74.30 (d'), 77.82 (d'), 80.93 (d'), 82.68 (d'), 105.43 (d'), 112.50 (s'); exact mass m/z calcd for $\text{C}_{11}\text{H}_{19}\text{O}_{10}\text{S}_2$ (M - CH_3) 375.0420, found 375.0419.

5,6-Dideoxy-1,2-O-isopropylidene-3-O-methyl- α -D-xylohex-5-enofuranose (17a) from Dimesylate 17. The general procedure was followed, but with some changes, using Te powder (200 mesh, 187.6 mg, 1.4703 mmol), Et_3BHLi (1 M in THF, 3.8 mL, 3.8 mmol), an initial reaction time of ca. 30 min at 75 °C (bath temperature), dimesylate **17** (573.4 mg, 1.4703 mmol) in THF (5 mL) (added at room temperature), and a reaction time of 12 h at room temperature. The mixture was washed out of the flask with Et₂O and evaporated. Flash chromatography of the residue over silica gel (2 \times 15 cm), using 1:1 EtOAc-hexane, gave **17a** (183 mg, 67%) as a clear oil, identical to an authentic⁶⁰ sample.

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(E)-3,4-Dideoxy-1,2:5,6-di-O-isopropylidene-erythro-3(E)-hex-3-enitol (18a) from Dimesylate 18. The general procedure was followed, but with minor changes, using Te powder (200 mesh, 191.1 mg, 1.4983 mmol), Et₃BHLi (1 M in THF, 3.9 mL, 3.9 mmol), an initial reaction time of ca. 30 min at 70 °C (bath temperature), 1,2:5,6-di-O-isopropylidene-3,4-bis-O-(methylsulfonyl)-D-glucitol⁶¹ (626.3 mg, 1.4983 mmol) in THF (5 mL) (injected over 2 h at room temperature), and a reaction time of 15 h. The mixture was washed out of the flask with CH₂Cl₂ and evaporated. Flash chromatography of the residue over silica gel (3 × 16 cm), using 3:7 EtOAc–hexane, gave **18a**^{24c,62} (254.5 mg, 75%) as a crystalline solid (needles): mp 70–71 °C (lit.^{24c,62} mp 69–71 °C).

(±)-2-exo-3-exo-Camphanediol 2,3-Bis(methanesulfonate) (21). MeSO₂Cl (0.39 mL, 5.0388 mmol) was added over 60 min to a stirred and cooled (ice bath) solution of (±)-2-exo-3-exo-camphane-2,3-diol⁶³ (170.0 mg, 1.00 mmol) and pyridine (0.65 mL, 8.0367 mmol) in CH₂Cl₂ (5 mL). The mixture was stirred at 0 °C for a further 10 min, the cold bath was removed, and stirring was continued for 5 days, at which point a major product was detected by TLC (silica, 10% EtOAc–CH₂Cl₂). The mixture was poured onto crushed ice (ca. 9 g) and washed successively with saturated aqueous NaHCO₃ (2 × 10 mL), aqueous CuSO₄ (10% w/v, 4 × 10 mL) and brine (10 mL). The organic solution was dried (MgSO₄) and evaporated. Flash chromatography of the residue over silica gel (1.5 × 15 cm), using 1:9 EtOAc–CH₂Cl₂, gave **21** (233.3 mg, 72%): mp 95–97 °C; FTIR (CHCl₃ cast) 2962, 1354, 1175, 1033, 963, 880 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ: 0.83 (s, 3 H), 1.00 (s, 3 H), 1.10–1.20 (m including s at δ 1.12, 5 H in all), 1.55–1.90 (m, 2 H), 2.12 (d, *J* = 5.3 Hz, 1 H), 3.09 (s, 6 H), 4.59 (d, *J* = 7.0 Hz, 1 H), 4.69 (d, *J* = 7.0 Hz, 1 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ 11.20 (q'), 20.49 (q'), 20.75 (q'), 23.38 (t'), 32.58 (t'), 38.56 (q'), 38.63 (q'), 47.72 (s'), 49.41 (s'), 50.72 (d'), 81.93 (d'), 85.62 (d'); exact mass *m/z* calcd for C₁₁H₁₈O₃S (M – CH₃SO₃H) 230.0977, found 230.0977.

Methyl 4,6-O-benzylidene-α-D-galactopyranoside 2,3-Bis(methanesulfonate) (22). MeSO₂Cl (0.20 mL, 2.583 mmol) was added over 20 min to a stirred and cooled (0 °C) solution of methyl 4,6-O-benzylidene-α-D-galactopyranoside⁶⁴ (140.2 mg, 0.497 mmol) and pyridine (0.35 mL, 4.327 mmol) in CH₂Cl₂ (3 mL). After a further 15 min, the ice bath was removed, and stirring was continued for 18 h. The mixture was poured onto crushed ice (ca. 5 g) and washed successively with 10% w/v aqueous CuSO₄ (until a constant blue color was maintained in the aqueous layer), saturated aqueous NaHCO₃ (2 × 10 mL), and brine. The organic extract was dried (MgSO₄) and evaporated. The resulting oil was triturated with Et₂O, and the dimesylate (201.1 mg, 92%) was obtained as a crystalline solid: mp 200–202 °C; FTIR (CH₂Cl₂, cast) 3036, 2940, 1173, 1076, 968, 955 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 3.0 (s, 3 H), 3.27 (s, 3 H), 3.44 (s, 3 H), 3.82–3.96 (m, 2 H), 4.05–4.15 (m, 1 H), 4.28–4.36 (m, 1 H), 4.90–4.95 (m including s at δ 4.94, 2 H in all), 5.05–5.09 (m, 1 H), 5.6 (s, 1 H), 7.35–7.50 (m, 5 H); ¹³C NMR (CD₂Cl₂, 75 MHz) δ 38.74 (q'), 38.82 (q'), 55.02 (q'), 64.20 (d'), 68.86 (t'), 75.67 (d'), 76.11 (d'), 78.38 (d'), 100.44 (d'), 102.46 (d'), 126.40 (d'), 128.71 (d'), 129.65 (d'), 137.30 (s'); exact mass *m/z* calcd. for C₁₆H₂₂O₁₀S₂ 438.0654, found 438.0644.

5'-O-(Triphenylmethyl)uridine 2',3'-Cyclic Sulfite (24). SOCl₂ (0.06 mL, 0.823 mmol) was added dropwise to a stirred and cooled (0 °C) solution of 5'-O-(triphenylmethyl)uridine (221.9 mg, 0.457 mmol) and pyridine (0.09 mL, 1.113 mmol) in CH₂Cl₂ (10 mL). After 15 min, the ice bath was removed, and stirring was continued for 24 h. The mixture was poured onto ice (ca. 15 g) and extracted with CH₂Cl₂ (1 × 5 mL). The organic extract was washed with saturated aqueous CuSO₄ (1 × 3 mL), saturated aqueous NaHCO₃ (1 × 2 mL), water (2

× 1 mL), and brine (1 × 2 mL), dried (MgSO₄), and evaporated. Flash chromatography of the residue over silica gel (2.5 × 15 cm), using 1:1 EtOAc–hexane, gave **24** (201.8 mg, 83%) as a colorless oil that consisted of a 2.5:1 mixture of diastereoisomers: FTIR (CH₂Cl₂ cast) 3202, 3060, 1694, 1219, 1003 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ (major isomer only) 3.47 (dd, *J* = 11.0, 3.5 Hz, 1 H), 3.54 (dd, *J* = 11.0, 5.0 Hz, 1 H), 4.34 (dd, *J* = 9.0, 4.0 Hz, 1 H), 5.56 (dd, *J* = 8.0, 2.0 Hz, 1 H), 5.65 (dd, *J* = 6.5, 4.0 Hz, 1 H), 5.69 (dd, *J* = 6.5, 2.5 Hz, 1 H), 5.87 (d, *J* = 2.5 Hz, 1 H), 7.25–7.50 (m, 16 H), 9.65 (br s, 1 H); ¹³C NMR (CDCl₃, 75.5 MHz) δ (major isomer only) 63.60 (t'), 84.84 (d', two overlapping signals), 87.31 (d'), 87.91 (s'), 92.15 (d'), 103.45 (d'), 127.80 (d'), 128.40 (d'), 128.98 (d'), 141.71 (d'), 143.61 (s'), 150.43 (s'), 163.42 (s'); exact mass *m/z* calcd for C₂₈H₂₄N₂O₅ (M – SO₂) 468.1685, found 468.1680.

(6S,7S)-1,11-Dodecadiene-6,7-diol (26). 4-Bromo-1-butene (678.3 mg, 5.0241 mmol) was added dropwise to a stirred suspension of Mg (174.2 mg, 7.1672 mmol) in Et₂O (2 mL) at such a rate that the mixture refluxed gently. The mixture was refluxed for 30 min after the end of the addition (oil bath at 40 °C) and then allowed to cool. The resulting Grignard reagent⁶⁵ was then added dropwise over 5 min by syringe to a stirred and cooled (–30 °C) suspension of anhydrous CuI (66.7 mg, 0.3502 mmol) in THF (5 mL). After a further 5 min, (*S,S*)-1,2,3,4-diepoxybutane⁶⁶ (115.8 mg, 1.3465 mmol) was added over 15 min. The cold bath was removed and replaced by an ice bath, and stirring was continued for 1.5 h. Saturated aqueous NH₄Cl was then added to the cold suspension, and the mixture was extracted with Et₂O (2 × 10 mL). The organic extract was dried (MgSO₄) and evaporated. Flash chromatography of the residue over silica gel (3 × 15 cm), using 2:3 EtOAc–hexane, gave **26** (208.1 mg, 91%) as a colorless oil: FTIR (CH₂Cl₂ cast) 3373, 3076, 2996 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ 1.33–1.65 (m, 8 H), 1.98–2.16 (m, 4 H), 2.25 (br s, *W*_{1/2} = 7.0 Hz, 2 H), 3.32–3.42 (m, 2 H), 4.91–5.07 (m, 4 H), 5.75–5.89 (m, 2 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 25.34, 33.40, 34.06, 74.65, 114.70, 139.18; exact mass *m/z* calcd for C₁₂H₂₂O₂ 198.1620, found 198.1620.

(6S,7S)-1,11-Dodecadienyl-6,7-diol Bis(methanesulfonate) (27). MeSO₂Cl (0.23 mL, 2.9703 mmol) was added over 15 min to a stirred and cooled (0 °C) solution of **26** (142.1 mg, 0.7177 mmol) and pyridine (0.29 mL, 3.5884 mmol) in CH₂Cl₂ (1.4 mL). The ice bath was removed, and stirring was continued for 6 h. The mixture was poured onto crushed ice (ca. 5 g) and washed successively with saturated aqueous CuSO₄ (2 × 8 mL), water (10 mL), saturated aqueous NaHCO₃ (2 × 10 mL), and brine. The organic extract was dried (MgSO₄) and evaporated. Flash chromatography of the residue over silica gel (3 × 20 cm), using 3:7 EtOAc–hexane, gave **27** (195.0 mg, 77%) as a yellow oil: FTIR (CH₂Cl₂ cast) 2940, 1356, 1174 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ 1.48–1.59 (m, 4 H), 1.63–1.87 (m, 4 H), 2.02–2.20 (m, 4 H), 3.06 (s, 6 H), 4.72–4.79 (m, 2 H), 4.95–5.07 (m, 4 H), 5.72–5.88 (m, 2 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 24.30 (t'), 29.99 (t'), 33.49 (t'), 39.07 (q'), 81.20 (d'), 115.49 (t'), 138.21 (d'); mass (CI) *m/z* calcd for C₁₄H₂₆O₆S₂ 372.2, found 372.2; exact mass *m/z* calcd for C₇H₁₂O₃S (M – C₇H₁₄O₃S) 176.0507, found 176.0507.

(6E)-1,6,11-Dodecatriene (28) from Dimesylate (27). The general procedure was followed, but with minor changes, using Te powder (89.6 mg, 0.7022 mmol), Et₃BHLi (1 M in THF, 1.46 mL, 1.46 mmol), an initial reaction time of ca. 60 min at 40 °C (bath temperature), dimesylate **27** (226.2 mg, 0.6381 mmol) in THF (1.3 mL) (added at room temperature), and a reaction time of 2 h. The dark brown suspension was then filtered through a small pad of Celite and evaporated. Kugelrohr distillation (25 °C, 0.1 mmHg) of the resulting yellow liquid gave **28**⁶⁷ (99.6 mg, 95%) as a colorless, volatile liquid, containing slight (<4%) impurities (¹H NMR, 300

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MHz): FTIR (CH₂Cl₂ cast) 3076, 2977, 1265 cm⁻¹; ¹H NMR (CD₂Cl₂, 300 MHz) δ 1.38–1.50 (m, 4 H), 1.96–2.10 (m, 8 H), 4.92–5.04 (m, 4 H), 5.39–5.44 (m, 2 H), 5.76–5.90 (m, 2 H); ¹³C NMR (CD₂Cl₂, 75.5 MHz) δ 29.34 (t'), 32.43 (t'), 33.64 (t'), 114.49 (t'), 130.74 (d'), 139.42 (d'); exact mass *m/z* calcd for C₁₂H₂₀ 164.1565, found 164.1568.

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amino)methylene]-5'-*O*-[bis(4-methoxyphenyl)phenylmethyl]adenosine. P. S. holds a Graduate Scholarship from CNPq (Brazil).

Supporting Information Available: NMR spectra for new compounds that were not analyzed (26 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

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